

# An introduction to R: part II

## The psych package

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Slides at <https://personality-project.org/r/apa/apa-short.pdf>

Additional slides <https://personality-project.org/r/apa/apa-short-2.pdf>)



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Descriptive and inferential statistics

Scores and Reliability

$\beta, \omega$  EFA

Mediation IRT multilevel FA and beyond

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## An introduction to the psych package



## Outline

## Part I: What is R, where did it come from, why use it

- Installing R and adding packages: the building blocks of R

## Part II: A brief introduction – an overview

- R is just a fancy (very fancy) calculator
  - Descriptive data analysis
  - Some inferential analysis

## Part III R is a powerful statistical system

- Data entry (detail and practice)
  - Descriptive (again)
  - Inferential (t and F with more practice)
  - Regression
  - Basic R commands

## Part IV: Psychometrics

- Reliability and its discontents
  - EFA CFA SEM

## Part V: Help and More Help

- List of useful commands

## Part VI: The psych package and more practice



## The psych package

1. Developed at NU over the past 12 years to make using R easier for psychologists
  2. Basically does the kind of statistics that my students and I find useful for personality, motivation and cognitive psychology
  3. Available at CRAN for PCs and Macs
  4. Development version (for Macs) is always available at the <http://personality-project.org/r> repository.
  5. Bugs are fixed and new versions with new toys are released about every 4-6 months.
  6. Version number reflects the year and month of release (1.7.5)
  7. Has several vignettes to describe what it does:
    - <http://personality-project.org/r/psych/vignettes/intro.pdf> An introduction
    - <http://personality-project.org/r/psych/vignettes/overview.pdf> An overview
    - [http://personality-project.org/r/psych/vignettes/psych\\_for\\_sem.pdf](http://personality-project.org/r/psych/vignettes/psych_for_sem.pdf) as a front end to doing sem



## Show all the functions in the psych package

```
objects("package:psych")
```

```
objects("package:psych")
[1] "%+%"                      "ability"                    "affect"                     "all.income"
[5] "alpha"                      "anova.psych"                "autoR"                      "Bechtoldt"
...
[49] "cohen.kappa"               "comorbidity"                "con2cat"                    "congeneric.s
[53] "cor.ci"                    "cor.plot"                  "cor.plot.upperLowerCi"    "cor.smooth"
...
[81] "cushny"                    "d2r"                       "densityBy"                 "describe"
...
[109] "epi.dictionary"            "equamax"                   "error.bars"                "error.bars.
...
[177] "ICC2latex"                 "iclust"                    "ICLUST"                    "ICLUST.clust
...
[201] "irt.fa"                    "irt.item.diff.rasch"       "irt.person.rasch"          "irt.response
...
[241] "mixed.cor"                 "mixedCor"                  "mlArrange"                 "mlPlot"
...
[253] "omega"                     "omega.diagram"              "omega.graph"                "omega2latex"
...
[309] "read.clipboard.upper"      "read.file"                 "read.file.csv"              "read.https"
...
[329] "score.alpha"                "score.irt"                  "score.irt.2"                "score.irt.p
[333] "score.items"                "score.multiple.choice"     "scoreFast"                  "scoreIrt"
...
[405] "Thurstone"                 "Thurstone.33"                "topBottom"                  "tr"
[409] "Tucker"                     "unidim"                    "varimin"                   "veg"
```



## Getting and cleaning data

### Get your data: using `read.file` or `read.clipboard`

From a website: define the file name

`R code`

```
fn <- "http://personality-project.org/r/datasets/Maps.mixx.mysql.epi.bf.txt"
fn #show it to check
[1] "http://personality-project.org/r/datasets/Maps.mixx.mysql.epi.bf.txt"
mydata <- read.file(fn, header=TRUE)
```

From a local file: find the file using `read.file`

`R code`

```
> my.data <- read.file() #will open a search window, read the file
#depending upon the suffix, will read .sav, .csv, .txt, .rds,
.rDa, etc.
```

From the clipboard: (first, go to the remote site, copy to the clipboard and then use the `read.clipboard` function).

`R code`

```
mydata <- read.clipboard() #or
mydata <- read.clipboard.tab() #if an excel file
my.data <- read.clipboard.csv() #if a tab delimited file
```



## Getting and cleaning data

### Checking the data using describe

```

> dim(mydata)
[1] 231 86
> describe(mydata)

vars n mean sd median trimmed mad min max range skew kurtosis se
id 1 231 66.82 45.13 58 64.14 50.41 1 160 159 0.45 -0.96 2.97
delighted 2 231 0.82 1.05 1 0.68 1.48 0 9 9 2.46 14.15 0.07
sociable 3 231 1.32 0.96 1 1.28 1.48 0 3 3 0.06 -1.03 0.06
jittery 4 231 0.55 0.78 0 0.39 0.00 0 3 3 1.37 1.23 0.05
hostile 5 231 0.35 0.85 0 0.17 0.00 0 9 9 5.34 45.21 0.06
sluggish 6 231 1.21 0.96 1 1.14 1.48 0 3 3 0.47 -0.70 0.06
depressed 7 231 0.56 0.83 0 0.39 0.00 0 3 3 1.45 1.37 0.05
...
ashamed 71 231 0.32 1.15 0 0.06 0.00 0 9 9 5.92 40.25 0.08
anxious 72 231 0.75 1.26 0 0.53 0.00 0 9 9 3.85 21.39 0.08
idle 73 231 0.98 1.15 1 0.83 1.48 0 9 9 3.11 18.20 0.08
epiE 74 231 13.33 4.14 14 13.49 4.45 1 22 21 -0.33 -0.06 0.27
epiS 75 231 7.58 2.69 8 7.77 2.97 0 13 13 -0.57 -0.02 0.18
epiImp 76 231 4.37 1.88 4 4.36 1.48 0 9 9 0.06 -0.62 0.12
epilie 77 231 2.38 1.50 2 2.27 1.48 0 7 7 0.66 0.24 0.10
...
traitanx 85 231 39.01 9.52 38 38.36 8.90 22 71 49 0.67 0.47 0.63
stateanx 86 231 39.85 11.48 38 38.92 10.38 21 79 58 0.72 -0.01 0.76

```



## Getting and cleaning data

## Cleaning the data using scrub

We want to change 9s in variables 2 - 73 into NA

```
> cleaned <- scrub(mydata, where=2:73, isvalue=9, newvalue=NA)
> describe(cleaned)
```

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis	se
id	1	231	66.82	45.13	58.0	64.14	50.41	1	160	159	0.45	-0.96	2.97
delighted	2	230	0.78	0.90	1.0	0.67	1.48	0	3	3	0.79	-0.52	0.06
sociable	3	231	1.32	0.96	1.0	1.28	1.48	0	3	3	0.06	-1.03	0.06
jittery	4	231	0.55	0.78	0.0	0.39	0.00	0	3	3	1.37	1.23	0.05
hostile	5	230	0.31	0.63	0.0	0.17	0.00	0	3	3	2.12	4.19	0.04
sluggish	6	231	1.21	0.96	1.0	1.14	1.48	0	3	3	0.47	-0.70	0.06
depressed	7	231	0.56	0.83	0.0	0.39	0.00	0	3	3	1.45	1.37	0.05
...													
ashamed	71	228	0.21	0.57	0.0	0.05	0.00	0	3	3	3.00	8.95	0.04
anxious	72	228	0.64	0.84	0.0	0.51	0.00	0	3	3	1.11	0.32	0.06
idle	73	229	0.91	0.88	1.0	0.82	1.48	0	3	3	0.64	-0.42	0.06
epiE	74	231	13.33	4.14	14.0	13.49	4.45	1	22	21	-0.33	-0.06	0.27
epiS	75	231	7.58	2.69	8.0	7.77	2.97	0	13	13	-0.57	-0.02	0.18
epiImp	76	231	4.37	1.88	4.0	4.36	1.48	0	9	9	0.06	-0.62	0.12
epilie	77	231	2.38	1.50	2.0	2.27	1.48	0	7	7	0.66	0.24	0.10
...													
traitanx	85	231	39.01	9.52	38.0	38.36	8.90	22	71	49	0.67	0.47	0.63
stateanx	86	231	39.85	11.48	38.0	38.92	10.38	21	79	58	0.72	-0.01	0.76





## Multiple ways to graphically display data

1. box.plots (Core R)
  2. Violin plots (`violinBy` in *psych* )
  3. Scatter Plot Matrix (SPLOM) plots (`pairs.panels` in *psych*)

First, lets just make a smaller data.frame and then issue two different graphic commands. \_\_\_\_\_

## R code

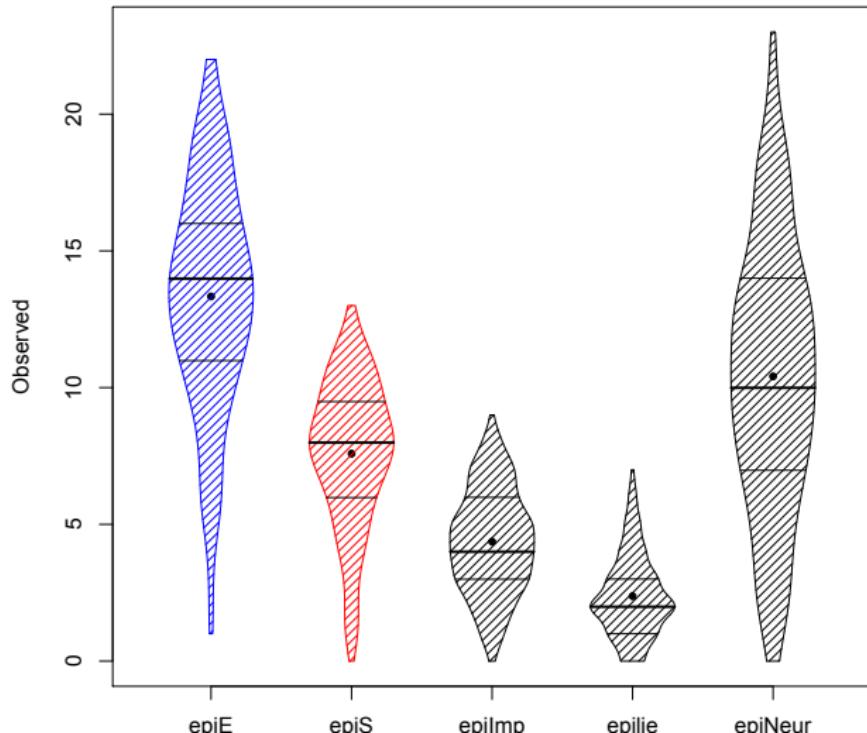
```
my.scales <- cleaned[74:86]
violinBy(my.scales[1:5])
pairs.panels(my.scales[c(1, 4, 5:10)], gap=0, pch=".")
```



## Graphical displays

```
Violin Plot violinBy(my.scales[1:5])
```

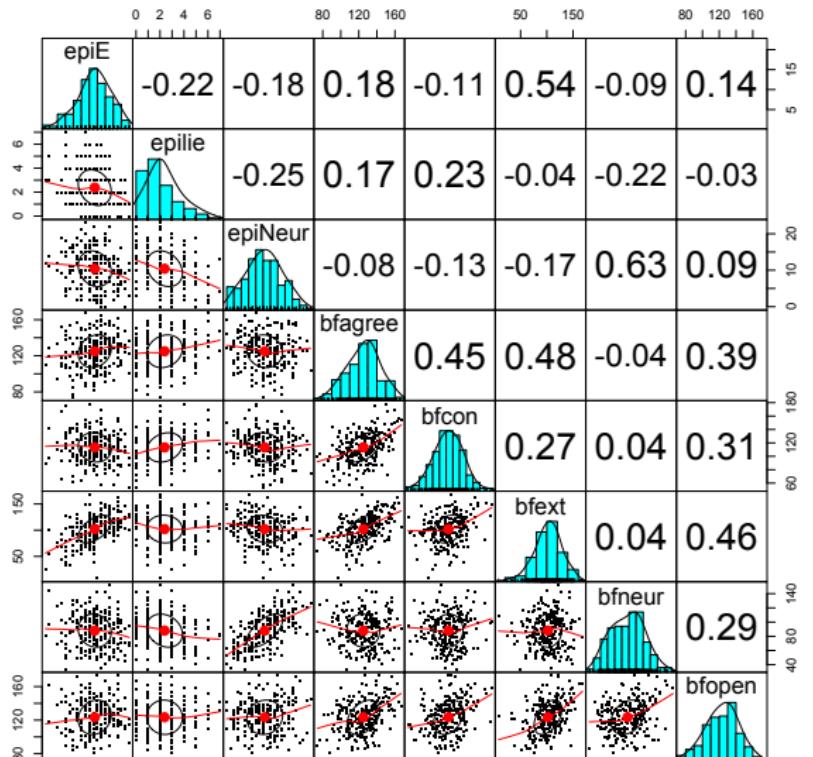
## Density plot



## Graphical displays

# Scatter Plot of Matrices (SPLOM) of select variables

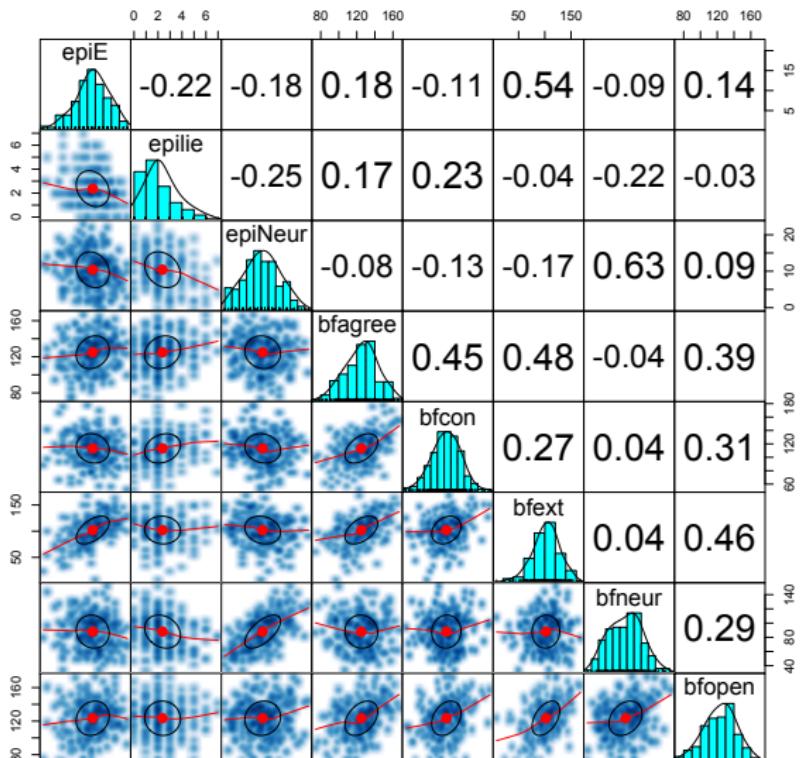
pairs.panels(my.scales[c(1,4,5:10)], gap=0, pch=". ")



## Graphical displays

## Scatter Plot of Matrices (SPLOM) of select variables

```
pairs.panels(my.scales[c(1,4,5:10)], gap=0, pch=". ", smoother=TRUE)
```





## Show a table of correlations

## R code

```
R <- lowerCor(my.scales[c(1, 4, 5:10)])
cor.plot(R, numbers=TRUE)
```

	epiE	epili	epiNr	bfagr	bfcon	bfext	bfner	bfopn
epiE	1.00							
epilie	-0.22	1.00						
epiNeur	-0.18	-0.25	1.00					
bfagree	0.18	0.17	-0.08	1.00				
bfcon	-0.11	0.23	-0.13	0.45	1.00			
bfext	0.54	-0.04	-0.17	0.48	0.27	1.00		
bfneur	-0.09	-0.22	0.63	-0.04	0.04	0.04	1.00	
bfopen	0.14	-0.03	0.09	0.39	0.31	0.46	0.29	1.00

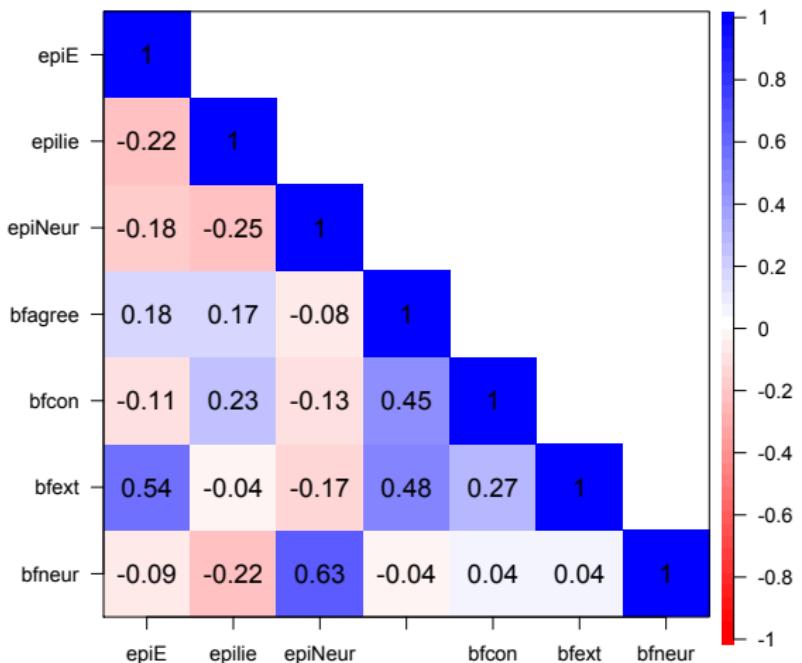
Automatically calls the `cor` and `round` functions with default parameters and then does a pretty print out using `lowerMat`. Invisibly returns the full (square) matrix of unrounded values.



## Graphical displays

## A simple heat map using cor.plot

## Correlation plot



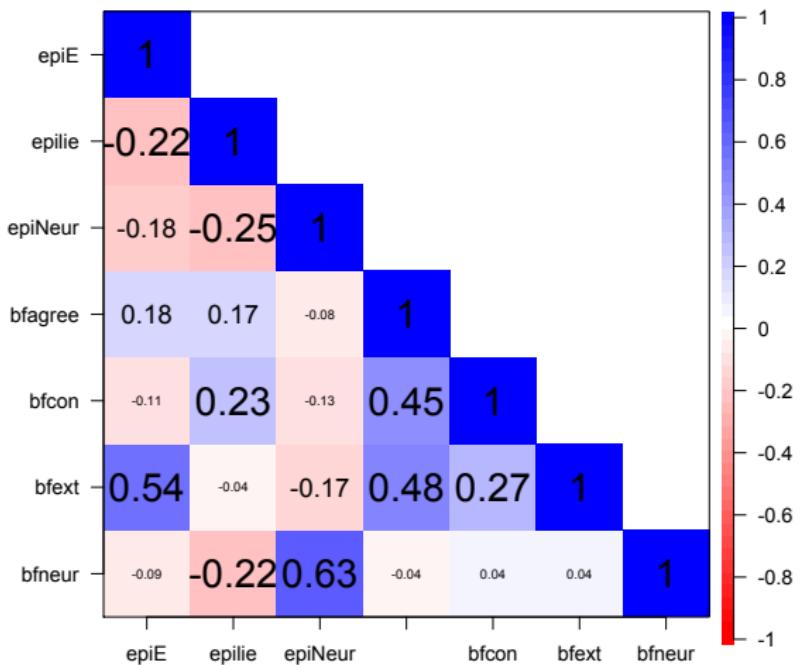
R code



## Graphical displays

## Scale the correlations using cor.plot

## Correlation plot



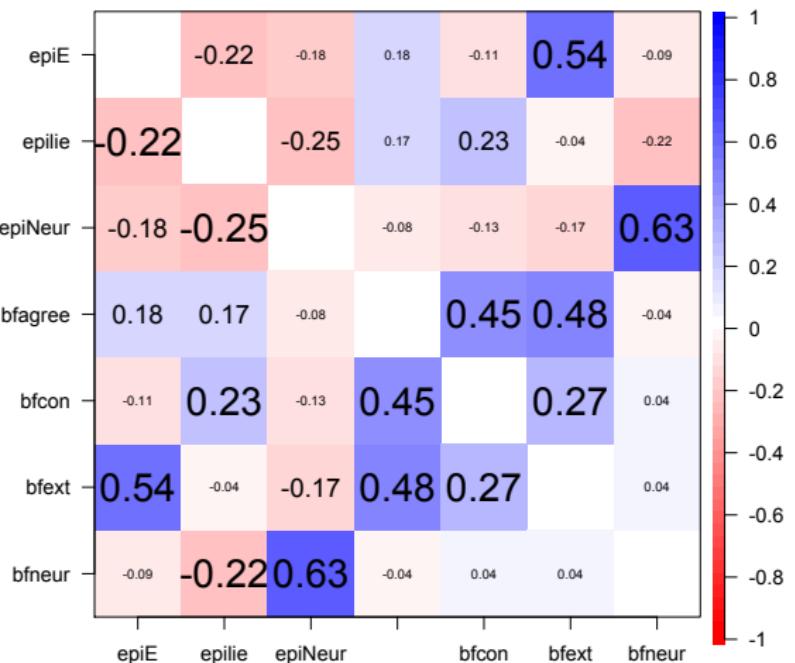
R code



## Graphical displays

## Show the whole matrix cor.plot

Correlation plot



R code  
`cor.plot(epi.bfi[c(1, 4:9)],  
numbers=TRUE, diag=FALSE)`



## Some inferential statistics – testing correlations

## Testing and displaying the “significance” of a set of correlations

1. Normal theory corr.test
    - raw probabilities as well as with a Holm adjusted for multiple correlations
  2. Display these with cor.plot
  3. Boot strapped confidence intervals based significance using cor.ci
    - Graphic displays correlations scaled by “significance”
    - Graphic displays of probability of correlation using plot.cor.upperLowerCi

## Code for the next slides

## R code

```
my.scales <- epi.bfi[c(1,4:10)]
corr.test(my.scales)
ci <- cor.ci(my.scales)
cor.plot.upperLowerCi(ci)
```



## Some inferential statistics – testing correlations

Normal theory test of correlations using corr.test

### R code

corr.test (mv.scales)

```

> corr.test(my.scales)
Call:corr.test(x = my.scales)
Correlation matrix
            epiE epilie epiNeur bfagree bfcon bfext bfneur bfopen
epiE      1.00 -0.22  -0.18    0.18 -0.11  0.54 -0.09  0.14
epilie   -0.22  1.00  -0.25    0.17  0.23 -0.04 -0.22 -0.03
epiNeur  -0.18 -0.25    1.00   -0.08 -0.13 -0.17  0.63  0.09
bfagree   0.18  0.17  -0.08    1.00  0.45  0.48 -0.04  0.39
bfcon    -0.11  0.23  -0.13    0.45  1.00  0.27  0.04  0.31
bfext     0.54 -0.04  -0.17    0.48  0.27  1.00  0.04  0.46
bfneur   -0.09 -0.22   0.63   -0.04  0.04  0.04  1.00  0.29
bfopen    0.14 -0.03   0.09    0.39  0.31  0.46  0.29  1.00

Sample Size
[1] 231

Probability values (Entries above the diagonal are adjusted for multiple tests.)
            epiE epilie epiNeur bfagree bfcon bfext bfneur bfopen
epiE      0.00  0.01  0.11   0.11  0.75  0.00  1.00  0.4
epilie   0.00  0.00  0.00   0.12  0.01  1.00  0.01  1.0
epiNeur  0.01  0.00  0.00   1.00  0.43  0.12  0.00  1.0
bfagree  0.01  0.01  0.21   0.00  0.00  0.00  1.00  0.0
bfcon   0.08  0.00  0.04   0.00  0.00  0.00  1.00  0.0
bfext    0.00  0.50  0.01   0.00  0.00  0.00  1.00  0.0
bfneur  0.15  0.00  0.00   0.50  0.50  0.57  0.00  0.0
bfopen  0.04  0.70  0.19   0.00  0.00  0.00  0.00  0.0

```

To see confidence intervals of the correlations, print with the short=FALSE option



A 2x8 grid of circles, arranged in two rows of eight circles each.

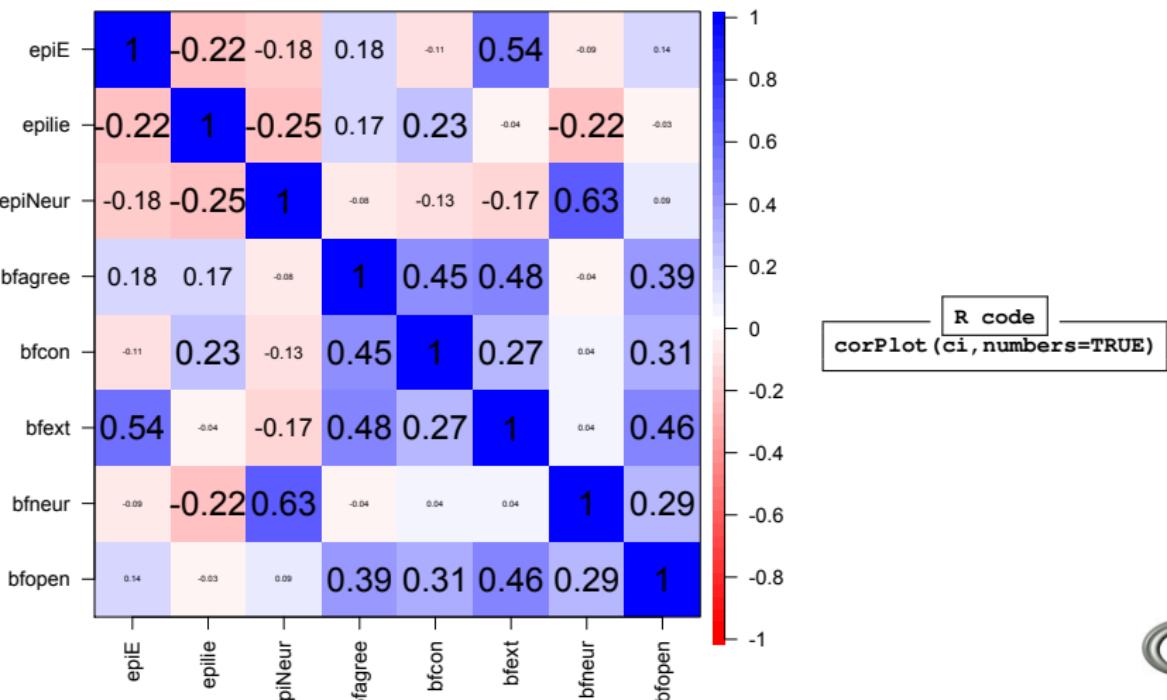
The diagram consists of four horizontal rows of circles. The top row has three circles. The second row has four circles. The third row has eight circles. The bottom row has four circles.

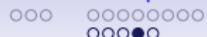
○○○○

## Some inferential statistics – testing correlations

Heat map scaled by “significance” using `cor.ci`

## Correlation plot

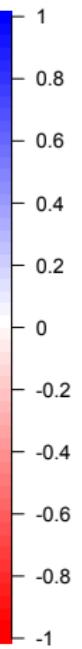
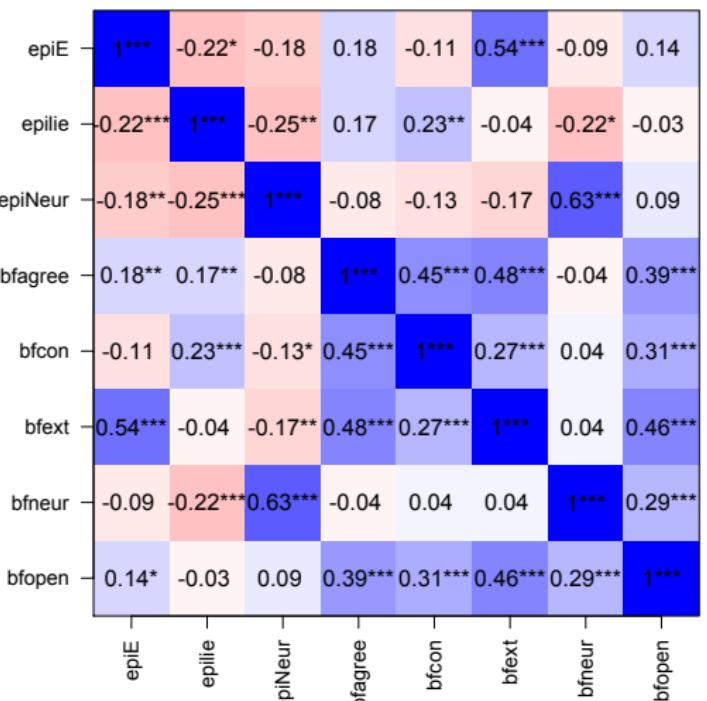




Some inferential statistics – testing correlations

## Heat map scaled by “significance” and showing magic asterisks

P values above the diagonal are Holm corrected



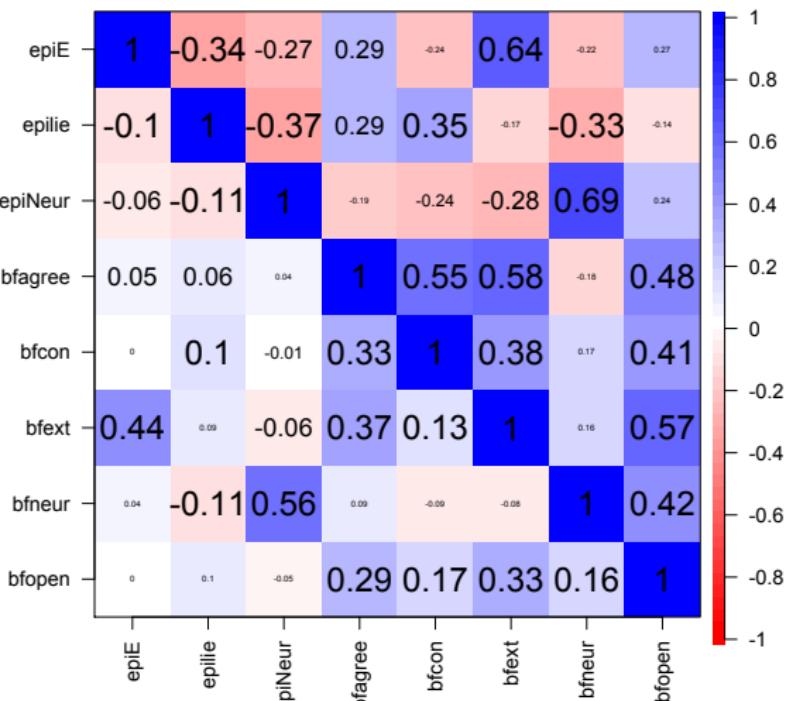
```
R code
corPlot(my.scales, stars=TRUE,
        numbers=TRUE, xlas=3,
        main="P values above the
        diagonal are Holm corrected")
```



Some inferential statistics – testing correlations

## Heat map scaled by “significance” and showing confidence intervals

Upper and lower confidence intervals of correlations



R code  
cor.plot.upperLowerCi(ci)



## Multiple types of reliability

### 1. Internal consistency estimates

- $\alpha, \lambda_6$ , use the alpha function
- $\omega_{hierarchical}$  and  $\omega_{total}$  use the omega function

### 2. IntraClass coefficients

- ICC

### 3. Rater agreement use wkappa function (finds Cohen's kappa and weighted kappa)

### 4. Multilevel reliability and generalizability, use mlr or multilevel.reliability



## Reliability

For the next examples we will use a built in data set

1. bfi consists of 25 personality items measuring 5 factors as well as some demographics.
  2. The data were collected as part of the SAPA project and have 2,800 subjects.
  3. For help on this data set, ?bfi
  4. To see all of the *psych* data sets: data(package="psych")



## Reliability

## First, we intentionally misspecify the data

R code

alpha(bfi[1:5]) #score the first five items

Some items ( A1 ) were negatively correlated with the total scale and probably should be reversed.

To do this, run the function again with the 'check.keys=TRUE' option  
Reliability analysis

Call: alpha(x = bfi[1:5])

```
raw_alpha std.alpha G6(smc) average_r S/N ase mean sd
 0.43      0.46      0.53      0.15 0.85 0.016 4.2 0.74
```

lower	alpha	upper	95% confidence boundaries
0.4	0.43	0.46	

Reliability if an item is dropped:

	raw_alpha	std.alpha	G6(smc)	average_r	S/N	alpha	se
A1	0.72	0.73	0.67	0.398	2.64	0.0087	
A2	0.28	0.30	0.39	0.097	0.43	0.0219	
A3	0.18	0.21	0.31	0.061	0.26	0.0249	
A4	0.25	0.31	0.44	0.099	0.44	0.0229	
A5	0.21	0.24	0.36	0.072	0.31	0.0238	

Item statistics

	n	raw.r	std.r	r.r	r.cor	r.drop	mean	sd	
A1	2784	0.066	0.024	-0.39	-0.31	2.4	1.4		<-- need to rekey this item
A2	2773	0.630	0.666	0.58	0.37	4.8	1.2		
A3	2774	0.724	0.742	0.72	0.48	4.6	1.3		
A4	2781	0.686	0.661	0.50	0.37	4.7	1.5		
A5	2784	0.700	0.719	0.64	0.45	4.6	1.3		

Non missing response frequency for each item



## Reliability

Try it again. Turn on automatic reversals. Get the scores

R code

```
scores <- alpha(bfi[1:5], check.keys =TRUE)
```

```
alpha(bfi[1:5], check.keys =TRUE)
```

Reliability analysis

```
Call: alpha(x = bfi[1:5], check.keys = TRUE)
```

raw_alpha	std.alpha	G6(smc)	average_r	S/N	ase	mean	sd
0.7	0.71	0.68	0.33	2.5	0.009	4.7	0.9

lower alpha upper      95% confidence boundaries  
0.69 0.7 0.72

Reliability if an item is dropped:

	raw_alpha	std.alpha	G6(smc)	average_r	S/N	alpha	se
A1-	0.72	0.73	0.67	0.40	2.6	0.0087	
A2	0.62	0.63	0.58	0.29	1.7	0.0119	
A3	0.60	0.61	0.56	0.28	1.6	0.0124	
A4	0.69	0.69	0.65	0.36	2.3	0.0098	
A5	0.64	0.66	0.61	0.32	1.9	0.0111	
...							

Warning message:

```
In alpha(bfi[1:5], check.keys = TRUE) :
```

Some items were negatively correlated with total scale and were automatically reversed.  
This is indicated by a negative sign for the variable name.



## R functions will return objects without necessarily telling you

1. The basic logic of R is that you can do lots of calculations, but you might not want all the output.
2. The output is there, to be processed by other functions if you want, but you probably don't want to see all of it unless you ask.
3. Thus, alpha returns the scores based upon the scales you asked for, but doesn't show them, because they are so many,
4. The str command tells you the structure of an object. The names will just list the names of the objects.



## Reliability

## names and str of alpha output

R code

```
names(scores)
str(scores)
```

```
names(scores)
[1] "total"           "alpha.drop"      "item.stats"      "response.freq" "keys"
[5] "scores"          "nvar"           "boot.ci"         "call"           "title"
[9] "boot"            "Unidim"          "Fit"             "sd"             "mean"
[13] "average_r"       "S/N"             "ase"             "sd"             "mean"
[17] "G6(smc)"        "average_r"      "S/N"             "sd"             "mean"
[21] "raw_alpha"       "std.alpha"       "G6(smc)"        "average_r"     "S/N"
[25] "alpha.drop"      "raw_alpha"       "std.alpha"       "G6(smc)"       "average_r"
[29] "item.stats"      "raw.r"           "std.r"           "r.cor"          "r.drop"
[33] "n"               "r.drop"          "n"               "r.cor"          "r.drop"
[37] "alpha.se"         "alpha.se"         "alpha.se"         "alpha.se"        "alpha.se"
[41] "alpha_se"         "alpha_se"         "alpha_se"         "alpha_se"        "alpha_se
```

\$ total : 'data.frame': 1 obs. of 8 variables:

..\$ raw\_alpha: num 0.703

..\$ std.alpha: num 0.713

..\$ G6(smc) : num 0.683

..\$ average\_r: num 0.332

..\$ S/N : num 2.48

..\$ ase : num 0.00895

..\$ mean : num 4.65

..\$ sd : num 0.898

\$ alpha.drop : 'data.frame': 5 obs. of 6 variables:

..\$ raw\_alpha: num [1:5] 0.719 0.617 0.6 0.686 0.643

..\$ std.alpha: num [1:5] 0.726 0.626 0.613 0.694 0.656

..\$ G6(smc) : num [1:5] 0.673 0.579 0.558 0.65 0.605

..\$ average\_r: num [1:5] 0.398 0.295 0.284 0.361 0.322

..\$ S/N : num [1:5] 2.64 1.67 1.58 2.26 1.9

..\$ alpha se : num [1:5] 0.00873 0.0119 0.01244 0.00983 0.01115

\$ item.stats : 'data.frame': 5 obs. of 7 variables:

..\$ n : num [1:5] 2784 2773 2774 2781 2784

..\$ raw.r : num [1:5] 0.581 0.728 0.76 0.654 0.687

..\$ std.r : num [1:5] 0.566 0.748 0.767 0.631 0.699

..\$ r.cor : num [1:5] 0.376 0.667 0.709 0.471 0.596

..\$ r.drop: num [1:5] 0.308 0.564 0.587 0.394 0.489



## One of the objects of alpha is the scores object

**R code**

```
describe(scores$scores)
```

But, since there are scores for all subjects, but just one score, this is not very interesting.

```
describe(scores$scores)
  vars      n   mean    sd median trimmed   mad   min   max range skew kurt
X1      1 2800 4.65 0.9     4.8     4.73 0.89     1     6      5 -0.76
>
```

Note that alpha has the option of doing cumulative scores (adding up items, or scoring in the unit of the items (the default).

**R code**

```
scores <- alpha(bfi[1:5], check.keys=TRUE, cumulative=TRUE)
#set the cumulative option to be true
describe(scores$scores)
```

```
describe(scores$scores)
  vars      n   mean    sd median trimmed   mad   min   max range skew kurt
X1      1 2800 23.08 4.54     24     23.43 4.45     5    30     25 -0.73
```



## Scoring Scales

## Perhaps a more useful case: scoring multiple scales using scoreItems

First, define the scoring keys, and then call scoreItems Use the msq data set

R code

```
keys <- make.keys(msq[1:75],list(
+ EA = c("active", "energetic", "vigorous", "wakeful", "wide.awake", "full.of.pep",
+       "lively", "-sleepy", "-tired", "-drowsy"),
+ TA =c("intense", "jittery", "fearful", "tense", "clutched.up", "-quiet", "-still",
+       "-placid", "-calm", "-at.rest"),
+ PA =c("active", "excited", "strong", "inspired", "determined", "attentive",
+       "interested", "enthusiastic", "proud", "alert"),
+ NAf =c("jittery", "nervous", "scared", "afraid", "guilty", "ashamed", "distressed",
+       "upset", "hostile", "irritable" ))
+ )
> msq.scores <- scoreItems(keys,msq[1:75])
> msq.scores
```

```
all: scoreItems(keys = keys, items = msq[1:75])
```

```
(Unstandardized) Alpha:
      EA   TA   PA   NAf
alpha 0.93 0.75 0.92 0.83
```

```
Standard errors of unstandardized Alpha:
      EA     TA     PA     NAf
ASE  0.004 0.0082 0.0044 0.0064
```

```
Average item correlation:
      EA     TA     PA     NAf
average.r 0.58 0.23 0.52 0.33
```



**Scoring Scales**

## Score multiple scales (continued)

**Standard errors of unstandardized Alpha:**

	EA	TA	PA	NAf
ASE	0.004	0.0082	0.0044	0.0064

**Average item correlation:**

	EA	TA	PA	NAf
average.r	0.58	0.23	0.52	0.33

**Guttman 6\* reliability:**

	EA	TA	PA	NAf
Lambda.6	0.96	0.82	0.93	0.88

**Signal/Noise based upon av.r :**

	EA	TA	PA	NAf
Signal/Noise	14	3	11	4.9

**Scale intercorrelations corrected for attenuation**

raw correlations below the diagonal, alpha on the diagonal  
 corrected correlations above the diagonal:

	EA	TA	PA	NAf
EA	0.932	0.29	0.870	-0.069
TA	0.238	0.75	0.226	0.710
PA	0.804	0.19	0.915	0.044
NAf	-0.061	0.56	0.039	0.831



## Scoring Scales

## More detailed item statistics

R code

```
print(msq.scores, short=FALSE)
```

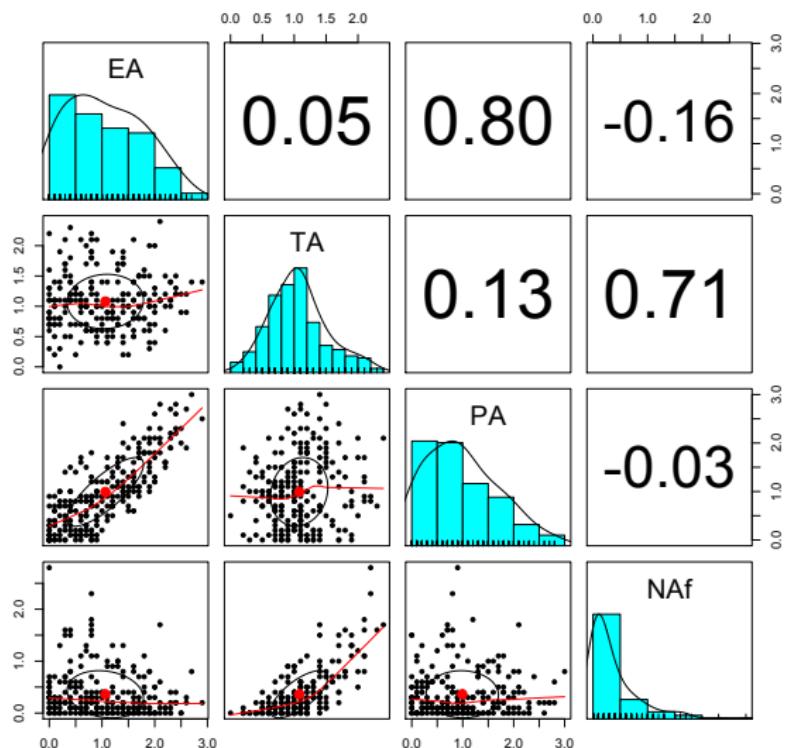
	Item by scale correlations:				Non missing response frequency for each item					
	corrected for item overlap and scale reliability				0	1	2	3	miss	
	EA	TA	PA	NAf	delighted	0.50	0.27	0.19	0.04	0.00
delighted	0.56	0.04	0.67	-0.17	sociable	0.24	0.30	0.35	0.11	0.00
sociable	0.57	0.06	0.64	-0.15	jittery	0.60	0.28	0.09	0.03	0.00
jittery	0.15	0.52	0.23	0.46	hostile	0.77	0.17	0.05	0.01	0.00
hostile	-0.21	0.37	-0.13	0.58	sluggish	0.24	0.44	0.19	0.13	0.00
sluggish	-0.65	-0.02	-0.40	0.21	depressed	0.61	0.26	0.08	0.05	0.00
depressed	-0.30	0.44	-0.26	0.67	satisfied	0.22	0.29	0.38	0.12	0.00
satisfied	0.54	-0.23	0.62	-0.35	relaxed	0.13	0.20	0.43	0.24	0.00
relaxed	0.35	-0.56	0.40	-0.51	warmhearted	0.17	0.23	0.37	0.22	0.00
warmhearted	0.47	-0.09	0.66	-0.19	blue	0.60	0.30	0.08	0.02	0.00
blue	-0.23	0.40	-0.19	0.64	intense	0.54	0.28	0.15	0.03	0.00
intense	0.25	0.42	0.44	0.47	strong	0.31	0.26	0.32	0.11	0.00
strong	0.55	0.00	0.69	-0.03	scared	0.80	0.15	0.04	0.01	0.00
scared	-0.05	0.61	0.08	0.75	enthusiastic	0.43	0.29	0.19	0.08	0.00
enthusiastic	0.69	0.16	0.83	-0.07	proud	0.41	0.24	0.26	0.10	0.00
proud	0.57	0.05	0.73	-0.11	sad	0.66	0.22	0.09	0.03	0.00
sad	-0.22	0.46	-0.18	0.73	active	0.38	0.32	0.22	0.08	0.00
active	0.71	0.18	0.83	-0.05	full.of.pep	0.54	0.21	0.20	0.05	0.00
full.of.pep	0.80	0.05	0.80	-0.15	unhappy	0.65	0.24	0.07	0.04	0.00
unhappy	-0.30	0.44	-0.26	0.70	lively	0.44	0.28	0.22	0.06	0.00
lively	0.78	0.04	0.81	-0.13						



## Scoring Scales

## Show the SPLOM of the msq scales using pairs.panels

Four msq scales



## Scoring Scales

## But what if we have overlapping scales?

1. Sometimes we are interested in how higher order scales relate to lower order scales.
2. The problem is, the items overlap.
3. Some people solve this problem by dropping the overlapping items. But this changes the meaning of the scales.
4. A fairly straightforward procedure is estimate the overlapping variances with the best estimate of shared (common) variance, similar to what is done when finding coefficient  $\alpha$ .
5. Need to do this on the correlation matrix of the items, not the raw data.
6. See ?scoreOverlap



## Scoring Scales

## Correcting for item overlap using scoreOverlap

R code

```

small.msq <- msq[ c("active", "energetic", "vigorous", "wakeful",
  "wide.awake", "full.of.pep", "lively", "sleepy", "tired", "drowsy",
  "intense", "jittery", "fearful", "tense", "clutched.up", "quiet",
  "still", "placid", "calm", "at.rest") ]
small.R <- cor(small.msq,use="pairwise")
keys.list <- list(
EA = c("active", "energetic", "vigorous", "wakeful", "wide.awake",
"full.of.pep", "lively", "-sleepy", "-tired", "-drowsy"),
TA =c("intense", "jittery", "fearful", "tense", "clutched.up",
"-quiet", "-still", "-placid", "-calm", "-at.rest"),
high.EA = c("active", "energetic", "vigorous", "wakeful",
"wide.awake", "full.of.pep", "lively"),
low.EA =c("sleepy", "tired", "drowsy"),
lowTA= c("quiet", "still", "placid", "calm", "at.rest"),
highTA = c("intense", "jittery", "fearful", "tense", "clutched.up")
)

keys <- make.keys(small.R,keys.list)

adjusted.scales <- scoreOverlap(keys.list,small.R)

```



## Scoring Scales

## Correcting for item overlap using scoreOverlap. (continued)

Call: scoreOverlap(keys = keys.list, r = small.R)

(Standardized) Alpha:

EA	TA	high.EA	low.EA	lowTA	highTA
0.93	0.75	0.94	0.93	0.73	0.76

(Standardized) G6\*:

EA	TA	high.EA	low.EA	lowTA	highTA
0.88	0.68	0.94	0.90	0.73	0.75

Average item correlation:

EA	TA	high.EA	low.EA	lowTA	highTA
0.59	0.23	0.68	0.81	0.35	0.38

Number of items:

EA	TA	high.EA	low.EA	lowTA	highTA
10	10	7	3	5	5

Signal to Noise ratio based upon average r and n

EA	TA	high.EA	low.EA	lowTA	highTA
14.1	3.0	14.8	12.9	2.7	3.1

Scale intercorrelations corrected for item overlap and attenuation  
adjusted for overlap correlations below the diagonal, alpha on the diagonal  
corrected correlations above the diagonal:

	EA	TA	high.EA	low.EA	lowTA	highTA
EA	0.93	0.27	0.965	-0.803	-0.18	0.253
TA	0.23	0.75	0.282	-0.167	-0.81	0.821
high.EA	0.90	0.24	0.937	-0.620	-0.12	0.324
low.EA	-0.75	-0.14	-0.579	0.928	0.25	-0.023
lowTA	-0.15	-0.60	-0.098	0.204	0.73	-0.335
highTA	0.21	0.10	0.070	0.010	0.25	0.777



## Scoring Scales

## Compare scoreOverlap with non-adjusted

R code

```
adjusted.scales <- scoreOverlap(keys.list, small.R)
raw <- scoreItems(keys.list, small.R)
```

Scale intercorrelations corrected for item overlap and attenuation  
 adjusted for overlap correlations below the diagonal, alpha on the diagonal  
 corrected correlations above the diagonal:

	EA	TA	high.EA	low.EA	lowTA	highTA
EA	0.93	0.27	0.965	-0.803	-0.18	0.253
TA	0.23	0.75	0.282	-0.167	-0.81	0.821
high.EA	0.90	0.24	0.937	-0.620	-0.12	0.324
low.EA	-0.75	-0.14	-0.579	0.928	0.25	-0.023
lowTA	-0.15	-0.60	-0.098	0.204	0.73	-0.335
highTA	0.21	0.62	0.273	-0.019	-0.25	0.757

Scale intercorrelations corrected for attenuation  
 raw correlations below the diagonal, alpha on the diagonal  
 corrected correlations above the diagonal:

	EA	TA	high.EA	low.EA	lowTA	highTA
EA	0.93	0.27	1.024	-0.848	-0.18	0.253
TA	0.23	0.75	0.282	-0.167	-1.06	1.056
high.EA	0.96	0.24	0.937	-0.620	-0.12	0.324
low.EA	-0.79	-0.14	-0.579	0.928	0.25	-0.023
lowTA	-0.15	-0.78	-0.098	0.204	0.73	-0.335
highTA	0.21	0.80	0.273	-0.019	-0.25	0.757



## $\alpha$ , $\omega_{hierarchical}$ and $\beta$ as alternative measures of internal consistency

### 1. $\alpha$ as the mean split half reliability

- alpha to find  $\alpha$
- splitHalf to find all (if  $n \leq 16$ ) or 10,000 random possible split half reliabilities ( $n > 16$ )

### 2. $\omega_{hierarchical}$ and $\omega_{total}$ as factor based reliabilities

- $\omega_{hierarchical}$  estimates general factor saturation
- Found using omega and omegaSem

### 3. $\beta$ as worst split half reliability as an alternative estimate of the general factor saturation.

- Found using a hierarchical clustering algorithm (iclust).
- iclust is also useful for scale construction.



## $\alpha$ from alpha and all split halves found using splitHalf

Find  $\alpha$  and all split half reliabilities of 5 Agreeableness items and 5 Conscientiousness items from the bfi data set included in psych.

R code

```
alpha(bfi[1:10]) #find alpha, let it automatically reverse items
splitHalf(bfi[1:10], keys=c(1, 9, 10)) #reverse 3 items
```

```
Reliability analysis
Call: alpha(x = bfi[1:10])
      raw_alpha std.alpha G6(smc) average_r S/N   ase mean     sd
      0.73       0.74      0.76      0.22 2.8 0.01  4.5 0.73
lower alpha upper      95% confidence boundaries
0.71 0.73 0.75
```

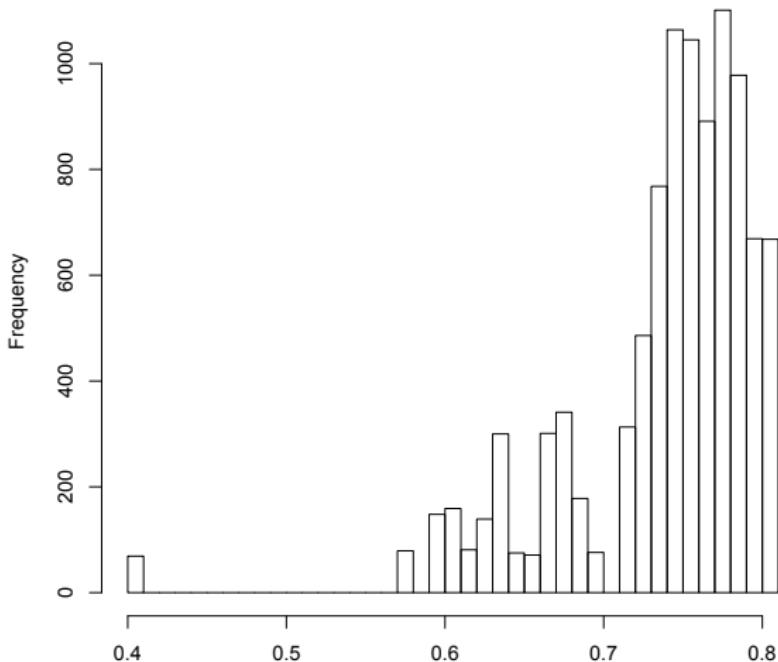
```
Split half reliabilities
Call: splitHalf(r = bfi[1:10], keys = c(1, 9, 10))
```

```
Maximum split half reliability (lambda 4) = 0.81
Guttman lambda 6 = 0.76
Average split half reliability = 0.73
Guttman lambda 3 (alpha) = 0.74
Minimum split half reliability (beta) = 0.41
```



All possible spit halves of 5 agreeableness and 5 conscientiousness items. Note the one worst one! This is not one construct.

All split half reliabilities of bfi[1:10]



Estimating  $\omega_{hierarchical}$  and  $\omega_{total}$  using omega

## Using the omega function

R code

`omega(ability, 4)`

Omega

Call: omega(m = ability, nfactors = 4)

Alpha:	0.83
G.6:	0.84
Omega Hierarchical:	0.65
Omega H asymptotic:	0.76
Omega Total	0.86

Schmid Leiman Factor loadings greater than 0.2

	g	F1*	F2*	F3*	F4*	h2	u2	p2
reason.4	0.50			0.27		0.34	0.66	0.73
reason.16	0.42			0.21		0.23	0.77	0.76
reason.17	0.55			0.47		0.52	0.48	0.57
reason.19	0.44			0.21		0.25	0.75	0.77
letter.7	0.52		0.35			0.39	0.61	0.69
letter.33	0.46		0.30			0.31	0.69	0.70
letter.34	0.54		0.38			0.43	0.57	0.67
letter.58	0.47		0.20			0.28	0.72	0.78
matrix.45	0.40				0.66	0.59	0.41	0.27
matrix.46	0.40				0.26	0.24	0.76	0.65
matrix.47	0.42					0.23	0.77	0.79
matrix.55	0.28					0.12	0.88	0.65
rotate.3	0.36	0.61				0.50	0.50	0.26
rotate.4	0.41	0.61				0.54	0.46	0.31
rotate.6	0.40	0.49				0.41	0.59	0.39
rotate.8	0.32	0.53				0.40	0.60	0.26

With eigenvalues of:

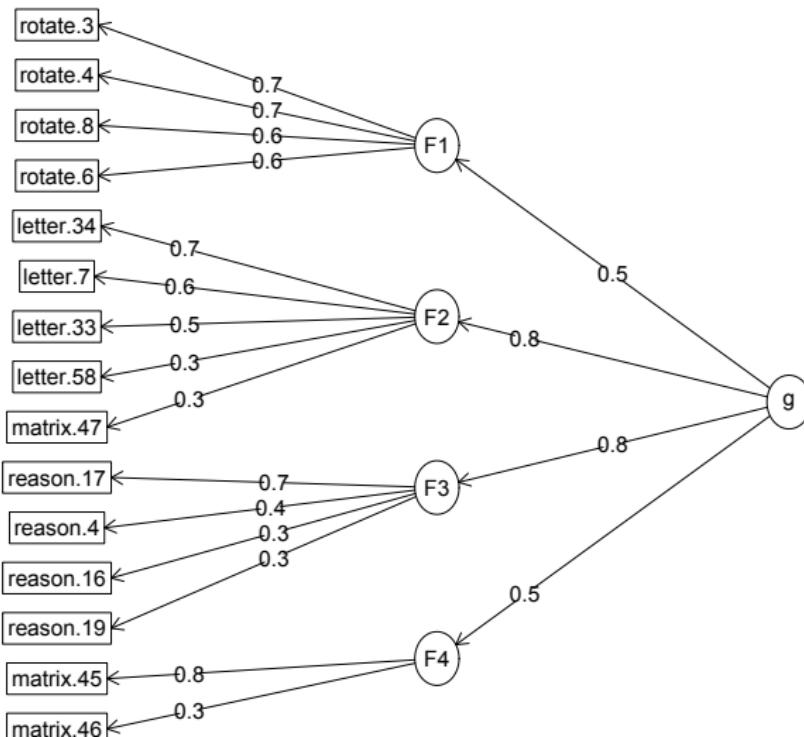
g F1\* F2\* F3\* F4\*



Estimating  $\omega_{hierarchical}$  and  $\omega_{total}$  using omega

# 16 ability items from the International Cognitive Ability Resource

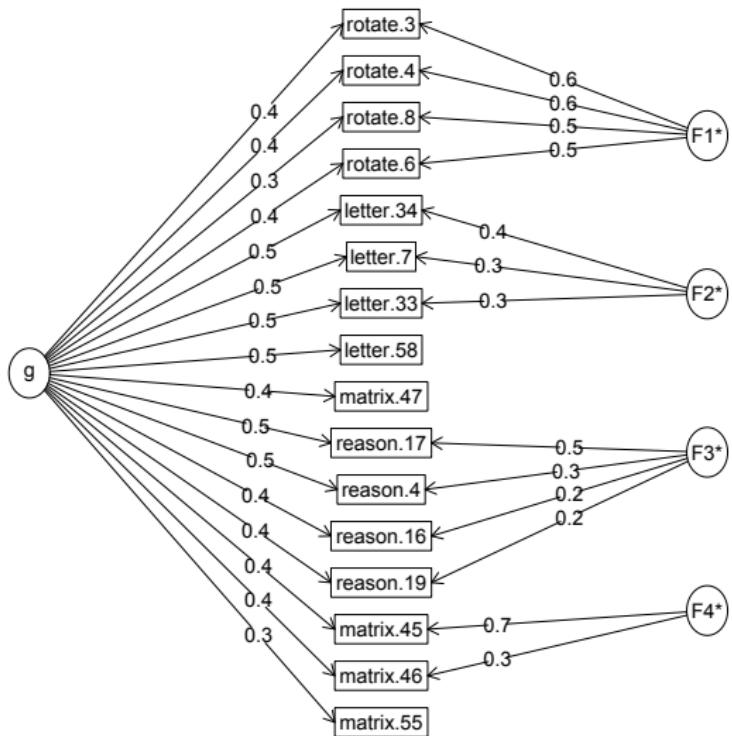
general ability and 4 subfactors of ICAR data



Estimating  $\omega_{hierarchical}$  and  $\omega_{total}$  using omega

## Schmid Leiman transformation of 16 ability items from ICAR

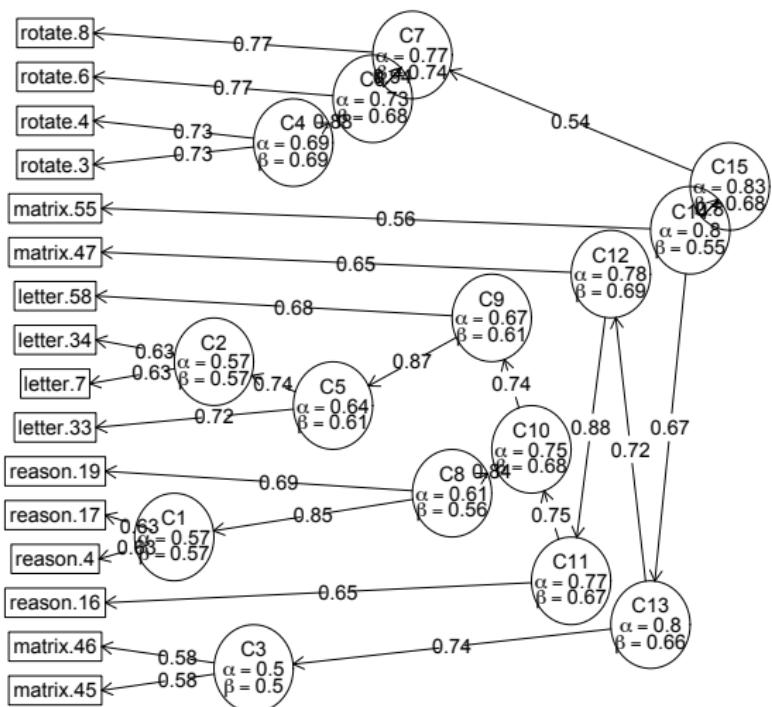
general ability and 4 subfactors of ICAR data



Estimating  $\omega_{hierarchical}$  and  $\omega_{total}$  using omega

## Hierarchical clustering of 16 ICAR ability items: iclust(ability)

Hierarchical clustering of 16 ability items using iclust



## Exploratory Factor Analysis

1. How many factors: an unsolved problem
  - Parallel analysis, MAPS, VSS, BIC, RMSEA, etc. available in `nfactors` and `fa.parallel`
2. Factor extraction algorithms available in the `fa` function
  - maximum likelihood, minimum residual, principal factor, ...
3. Factor rotation procedures are done using `GPArotation` package
  - orthogonal: varimax, quartimax, bifactor, ...
  - oblique: oblimin, geomin, biquartimin, ...
4. Displaying the solutions using `fa.plot`

Note, that EFA is not the same as Principal Components Analysis and the two should not be confused.

1. PCA done using `principal`



How many factors are in the mood data

The number of factors problem is easy and hard

## No best rule, one worst rule

"Solving the number of factors problem is easy, I do it everyday before breakfast. But knowing the right solution is harder." (Henry Kaiser)

1. Parallel analysis (Extract factors until the eigen values are less than those of a random matrix).
    - Although a good rule for 100-500 subjects, this will not do as well with many ( $>1000$ ) subjects.
  2. Velicer's Minimum Average Partial (MAP) is pretty good
  3. For items, the Very Simple Structure (VSS) criterion is pretty good.
  4. Multiple statistical tests, many have problems with sample size.
    - If you want few factors, run few subjects
    - If you want many factors, run many subjects
  5. One worst rule is the eigen value of 1.0 rule.



How many factors are in the mood data

## What about parallel analysis? Pearson R or polychoric $\rho$ ?

Of the raw (Pearson) correlations compared to the polychoric correlations

R code

```
> fa.parallel(cleaned[2:73])
Parallel analysis suggests that the number of factors = 5
and the number of components = 5
```

```
#use polychoric correlations
fa.parallel(cleaned[2:73],cor="poly")

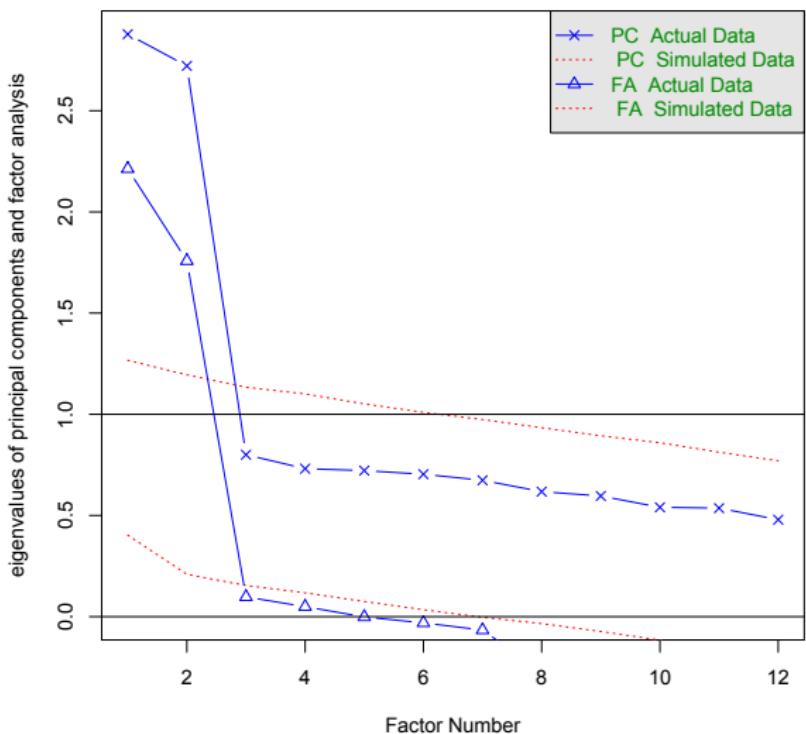
> fa.parallel(cleaned[2:73],cor="poly")
some warnings are issued
The items do not have an equal number of response
alternatives, global set to FALSE
Parallel analysis suggests that the number of factors = 4
and the number of components = 4
Warning message:
In cor.smooth(mat) : Matrix was not positive definite,
smoothing was done
>
```



How many factors are in the mood data

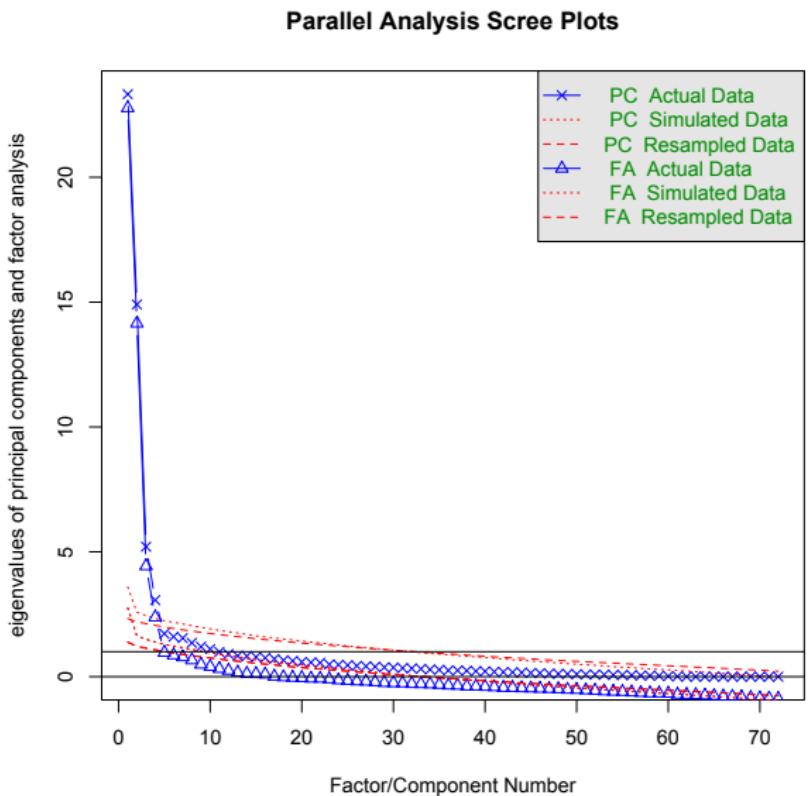
## Parallel analysis with Pearson correlations

Parallel Analysis Scree Plots



How many factors are in the mood data

## Parallel analysis with polychoric correlations (takes somewhat longer)



How many factors are in the mood data

## How many factors: what does nfactors tell us?

```
> nfactors(cleaned[2:73])
```

### Number of factors

```
Call: vss(x = x, n = n, rotate = rotate, diagonal = diagonal, fm = fm,
      n.obs = n.obs, plot = FALSE, title = title, use = use, cor = cor)
```

VSS complexity 1 achieves a maximum of 0.74 with 2 factors

VSS complexity 2 achieves a maximum of 0.91 with 3 factors

The Velicer MAP achieves a minimum of 0.01 with 9 factors

Empirical BIC achieves a minimum of -10081.25 with 6 factors

Sample Size adjusted BIC achieves a minimum of -1843.62 with 11 factors

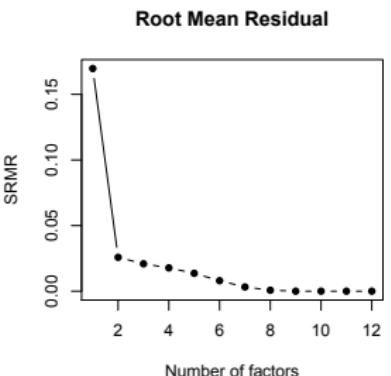
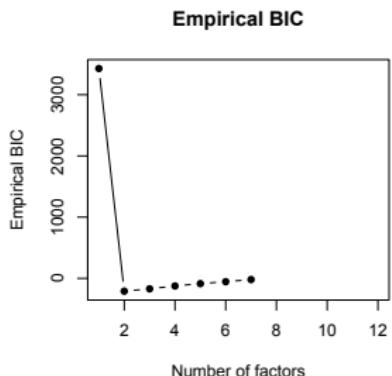
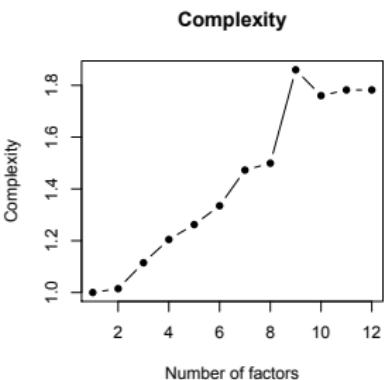
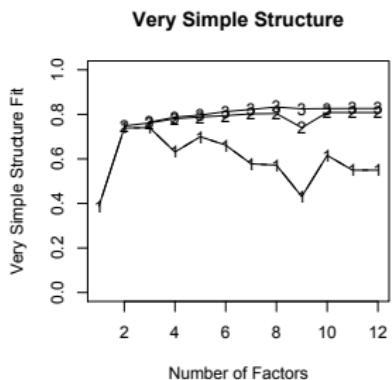
### Statistics by number of factors

	vssl	vss2	map	dof	chisq	prob	sqresid	fit	RMSEA	BIC	SABIC	complex	eChisq	SRMR	etc
1	0.68	0.00	0.0552	2484	8907	0.0e+00	192.3	0.68	0.115	-4612	3261	1.0	35253	0.173	0
2	0.74	0.90	0.0179	2413	5827	2.9e-282	58.9	0.90	0.087	-7305	343	1.2	7619	0.080	0
3	0.69	0.91	0.0145	2343	4928	8.6e-186	40.3	0.93	0.077	-7823	-397	1.4	4442	0.061	0
4	0.66	0.86	0.0110	2274	4095	1.7e-107	29.0	0.95	0.067	-8281	-1074	1.5	2532	0.046	0
5	0.62	0.85	0.0101	2206	3587	1.9e-69	24.9	0.96	0.061	-8419	-1427	1.7	1933	0.040	0
6	0.63	0.84	0.0094	2139	3274	4.1e-51	22.0	0.96	0.057	-8367	-1588	1.8	1560	0.036	0
7	0.63	0.83	0.0094	2073	3047	2.1e-40	19.7	0.97	0.055	-8235	-1665	1.9	1314	0.033	0
8	0.63	0.82	0.0091	2008	2810	7.0e-30	17.9	0.97	0.052	-8118	-1754	2.0	1100	0.031	0
...															
18	0.53	0.78	0.0105	1413	1467	1.5e-01	9.2	0.98	0.032	-6223	-1745	2.6	324	0.017	0
19	0.52	0.78	0.0108	1359	1371	4.0e-01	8.7	0.99	0.030	-6025	-1718	2.6	291	0.016	0
20	0.52	0.77	0.0111	1306	1284	6.6e-01	8.2	0.99	0.028	-5824	-1685	2.7	258	0.015	0



How many factors are in the mood data

## The number of factors from nfactors



How many factors are in the mood data

## What if we use polychoric correlations

```
> nfactors(cleaned[2:73], cor="poly")
The items do not have an equal number of response alternatives, global set to FALSE
```

Number of factors

```
Call: vss(x = x, n = n, rotate = rotate, diagonal = diagonal, fm = fm,
n.obs = n.obs, plot = FALSE, title = title, use = use, cor = cor)
```

VSS complexity 1 achieves a maximum of 0.76 with 2 factors

VSS complexity 2 achieves a maximum of 0.93 with 2 factors

The Velicer MAP achieves a minimum of 0.02 with 9 factors

Empirical BIC achieves a minimum of -9764.2 with 5 factors

Sample Size adjusted BIC achieves a minimum of 39801.49 with 20 factors

Statistics by number of factors

	vss1	vss2	map	dof	chisq	prob	sqresid	fit	RMSEA	BIC	SABIC	complex	eChisq	SRMR	eCRMS
1	0.66	0.00	0.099	2484	56503	0	280.2	0.66	0.33	42984	50857	1.0	56472	0.219	0.222
2	0.76	0.93	0.027	2413	51500	0	58.1	0.93	0.32	38368	46016	1.3	9088	0.088	0.090
3	0.70	0.92	0.023	2343	50169	0	31.0	0.96	0.32	37417	44843	1.4	4295	0.060	0.063
4	0.67	0.89	0.018	2274	48889	0	21.6	0.97	0.32	36513	43721	1.5	2649	0.047	0.050
5	0.67	0.89	0.018	2206	48230	0	18.6	0.98	0.32	36224	43216	1.7	2242	0.044	0.047
6	0.66	0.87	0.018	2139	47695	0	16.0	0.98	0.33	36053	42833	1.8	1930	0.040	0.044
7	0.64	0.86	0.017	2073	47098	0	13.7	0.98	0.33	35816	42386	1.9	1612	0.037	0.041
8	0.65	0.84	0.017	2008	46595	0	11.8	0.99	0.33	35666	42031	1.9	1391	0.034	0.039
...															
19	0.59	0.82	0.022	1359	43055	0	3.5	1.00	0.40	35659	39966	2.6	462	0.020	0.027
20	0.56	0.82	0.022	1306	42770	0	3.1	1.00	0.41	35662	39801	2.6	419	0.019	0.026

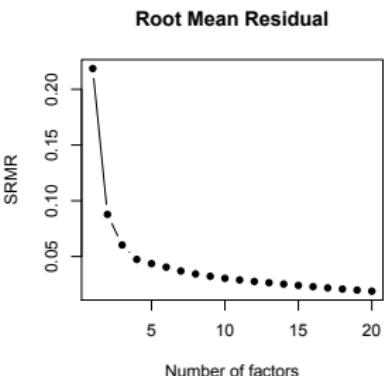
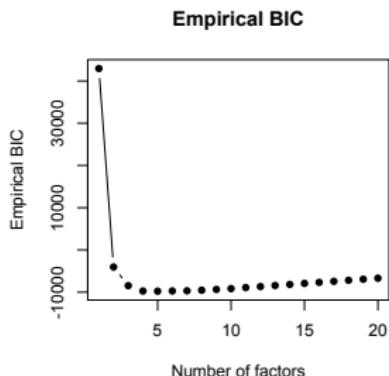
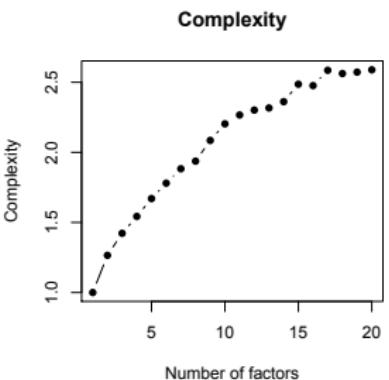
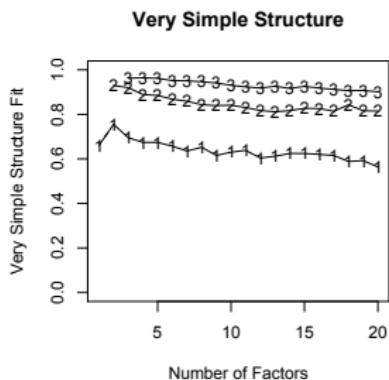
Warning message:

In cor.smooth(mat) : Matrix was not positive definite, smoothing was done



How many factors are in the mood data

## The number of factors from nfactors



## Factor extraction and graphical displays

## EFA of the Motivational State Questionnaire

## R code

```
f2 <- fa(msq[1:72], 2)  
summary(f2)
```

summary (f2)

Factor analysis with Call: fa(r = msg[1:72], nfactors = 2)

Test of the hypothesis that 2 factors are sufficient.

The degrees of freedom for the model is 2413 and the objective function value is -67730.13  
The number of observations was 3896 with Chi Square = 67730.13

The root mean square of the residuals (RMSA) is 0.09

The df corrected root mean square of the residuals is 0.09

Tucker Lewis Index of factoring reliability = 0.637

RMSEA index = 0.083 and the 90 % confidence intervals are 0.083

BTC = 47780.16

With factor correlations of

MR 1      MR 2

MB1 1 00 -0 13

MR2 -0.13 1.00



## Factor extraction and graphical displays

### Show the factors, sorted by factor loadings

```
> print(f2, sort=TRUE)
Factor Analysis using method = minres
Call: fa(r = msq[1:72], nfactors = 2, cor = "poly")
Standardized loadings (pattern matrix) based upon correlation matrix
      item   MR1   MR2   h2   u2 com
lively     20  0.89 -0.05 0.811 0.19 1.0
energetic   55  0.89  0.05 0.789 0.21 1.0
full.of.pep 18  0.89 -0.05 0.800 0.20 1.0
...
sluggish     5 -0.52  0.22 0.348 0.65 1.4
sleepy      59 -0.48  0.15 0.274 0.73 1.2
tired       28 -0.45  0.23 0.285 0.71 1.5
drowsy      51 -0.40  0.13 0.189 0.81 1.2
...
tense       69  0.14  0.85 0.714 0.29 1.1
frustrated   65 -0.10  0.83 0.718 0.28 1.0
ashamed      70  0.12  0.83 0.676 0.32 1.0
upset        48 -0.13  0.82 0.714 0.29 1.1
...
relaxed      8  0.44 -0.52 0.519 0.48 1.9
calm        50  0.26 -0.50 0.354 0.65 1.5
at.rest     26  0.38 -0.43 0.378 0.62 2.0
...
      MR1   MR2
SS loadings  21.07 17.15
Proportion Var 0.29  0.24
Cumulative Var 0.29  0.53
Proportion Explained 0.55  0.45
Cumulative Proportion 0.55  1.00
```



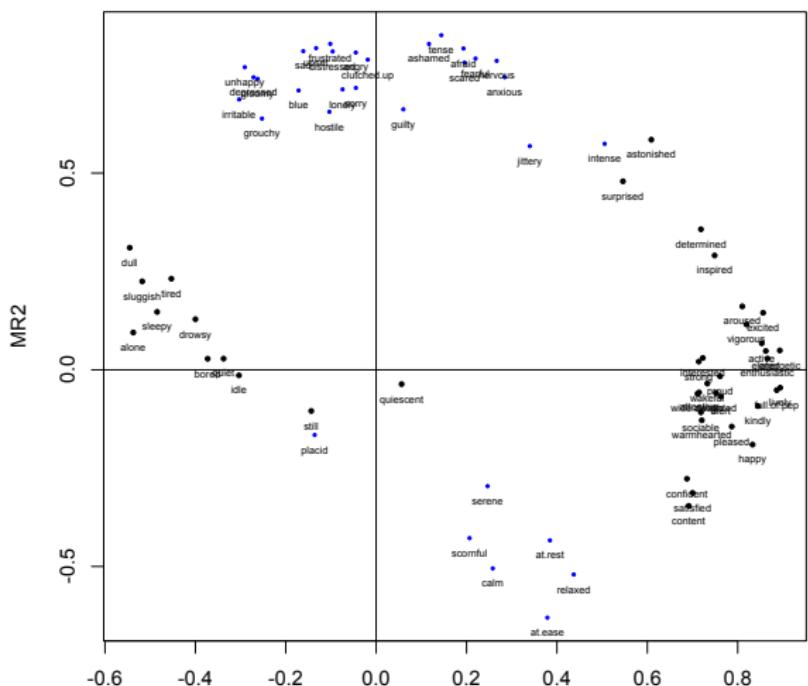
V <- Descriptive and inferential statistics  
ooo oooooooo  
ooo

## Mediation IRT multilevel FA and beyond

## Factor extraction and graphical displays

```
fa.plot(f2,labels=colnames(msq[1:72]),cex=.5,title="2 dimensions of  
the Motivational State Questionnaire")
```

## 2 dimensions of the Motivational State Questionnaire



Factor extraction and graphical displays

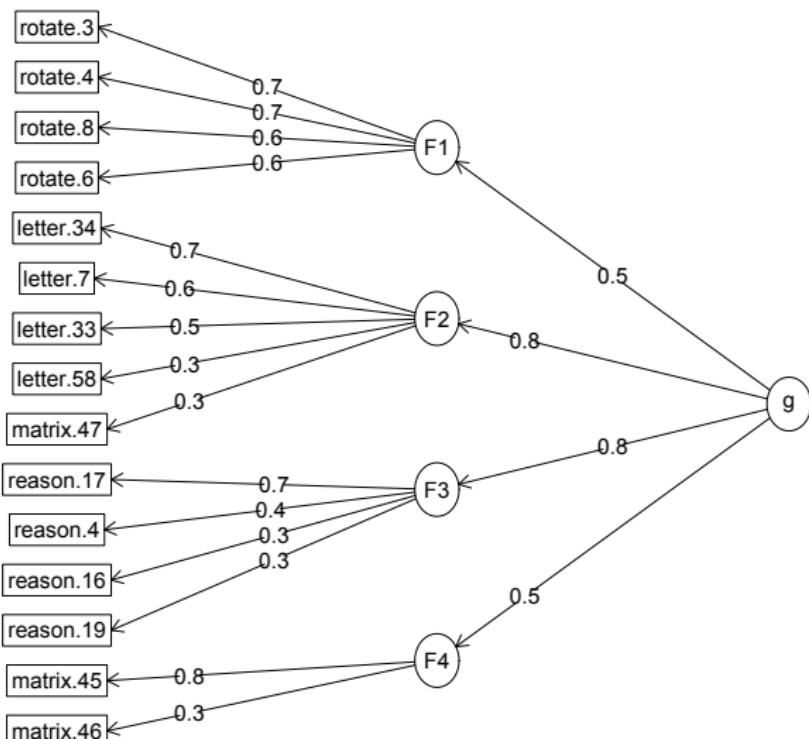
## Convert to and sort polar coordinates round(polar(f2),2)

	Var	theta21	vec121
<b>strong</b>	12	1.68	0.51
<b>enthusiastic</b>	14	1.89	0.75
...			
<b>anxious</b>	71	69.09	0.63
<b>nervous</b>	45	71.26	0.69
...			
<b>angry</b>	44	93.20	0.65
<b>sorry</b>	58	93.59	0.52
...			
<b>sad</b>	16	101.25	0.68
<b>blue</b>	10	103.59	0.53
...			
<b>drowsy</b>	51	162.17	0.18
<b>sleepy</b>	59	163.11	0.26
...			
<b>relaxed</b>	8	310.03	0.46
<b>at.rest</b>	26	311.56	0.34
..			
<b>happy</b>	61	347.14	0.73
<b>pleased</b>	60	349.61	0.64
...			
<b>alert</b>	52	354.89	0.59



## Graphical displays of hierarchical analysis

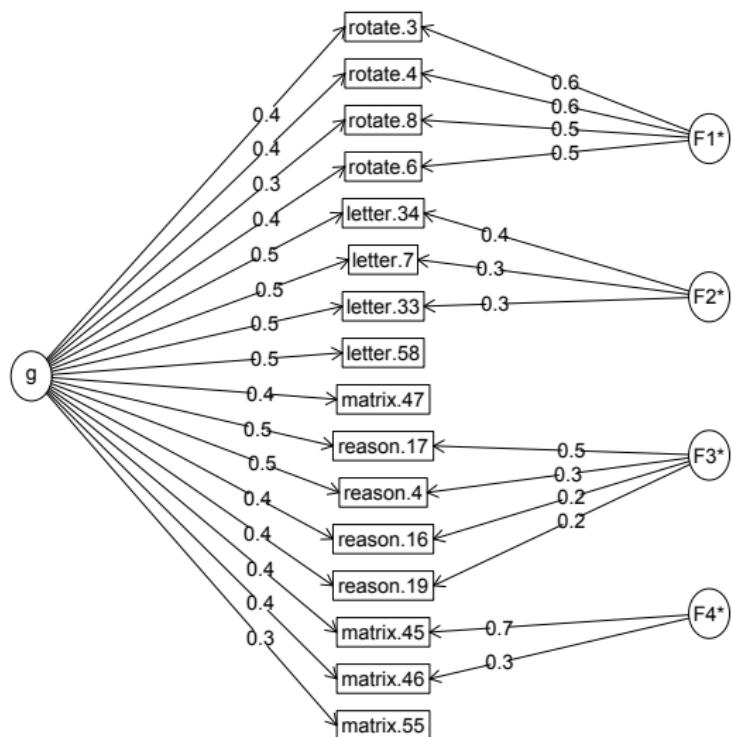
## 16 ability items from the International Cognitive Ability Resource general ability and 4 subfactors of ICAR data



## Graphical displays of hierarchical analysis

# Schmid Leiman transformation of 16 ability items from ICAR

general ability and 4 subfactors of ICAR data



## Graphical displays of hierarchical analysis

## More types of reliability

1.  $\alpha$  is a hodgepodge ratio of general factor and group factor reliability
  2.  $\omega_h$  (omega hierarchical) is an estimate of the general factor variance of a test
  3.  $\omega_t$  (omega total) is an estimate of the total reliable variance of a test
  4. When do we use these?
    - When estimating how much of a test measures one thing.  
 $\omega_{\alpha_h}$
    - When estimating what is the total reliable variance in a test  
(when adjusting for test reliability in an SEM context)



## Graphical displays of hierarchical analysis

## $\omega_h$ and $\omega_f$ reliabilities

## R code

```
om <- omega(ability, nfactors=4)
```

```

om
Omega
Call: omega(m = ability, nfactors = 4)
Alpha:          0.83
G.6:           0.84
Omega Hierarchical: 0.65
Omega H asymptotic: 0.76
Omega Total      0.86

Schmid Leiman Factor loadings greater than 0.2
      g   F1*   F2*   F3*   F4*   h2   u2   p2
reason.4  0.50           0.27       0.34  0.66  0.73
reason.16  0.42           0.21       0.23  0.77  0.76
reason.17  0.55           0.47       0.52  0.48  0.57
reason.19  0.44           0.21       0.25  0.75  0.77
letter.7   0.52           0.35       0.39  0.61  0.69
letter.33  0.46           0.30       0.31  0.69  0.70
letter.34  0.54           0.38       0.43  0.57  0.66
letter.58  0.47           0.20       0.28  0.72  0.78
matrix.45  0.40           0.66       0.66  0.59  0.41  0.27
matrix.46  0.40           0.26       0.26  0.24  0.76  0.65
matrix.47  0.42           0.23       0.23  0.77  0.79
matrix.55  0.28           0.12       0.12  0.88  0.65
rotate.3   0.36  0.61     0.50       0.50  0.50  0.26
rotate.4   0.41  0.61     0.54       0.46  0.46  0.31
rotate.6   0.40  0.49     0.41       0.59  0.59  0.39
rotate.8   0.32  0.53     0.40       0.60  0.60  0.26

```



## Graphical displays of hierarchical analysis

### $\omega$ continued

With eigenvalues of:

g	F1*	F2*	F3*	F4*
3.04	1.32	0.46	0.42	0.55

general/max 2.3 max/min = 3.17

mean percent general = 0.58 with sd = 0.2 and cv of 0.35

Explained Common Variance of the general factor = 0.53

The degrees of freedom are 62 and the fit is 0.05

The number of observations was 1525 with Chi Square = 70.19 with prob < 0.22

The root mean square of the residuals is 0.01

The df corrected root mean square of the residuals is 0.02

RMSEA index = 0.009 and the 90 % confidence intervals are 0 0.014

BIC = -384.25

Compare this with the adequacy of just a general factor and no group factors

The degrees of freedom for just the general factor are 104 and the fit is 0.78

The number of observations was 1525 with Chi Square = 1186.18 with prob < 5e-183

The root mean square of the residuals is 0.09

The df corrected root mean square of the residuals is 0.09

RMSEA index = 0.083 and the 90 % confidence intervals are 0.078 0.085

BIC = 423.88

#### Measures of factor score adequacy

	g	F1*	F2*	F3*	F4*
Correlation of scores with factors		0.83	0.80	0.53	0.56 0.71
Multiple R square of scores with factors		0.69	0.64	0.28	0.32 0.50
Minimum correlation of factor score estimates	0.37	0.28	-0.44	-0.37	0.00

Total, General and Subset omega for each subset



## Data set from Preacher & Hayes (2004)

R code

```
# from Preacher and Hayes (2004)
sobel <- structure(list(SATIS = c(-0.59, 1.3, 0.02, 0.01, 0.79, -0.35,
-0.03, 1.75, -0.8, -1.2, -1.27, 0.7, -1.59, 0.68, -0.39, 1.33,
...
"Therapy", "Attributional Positivity"), .Names = c("SATIS", "THERAPY",
"ATTRIB")))
#n.iter set to 50 (instead of default of 5000) for speed of example
mediate(1,2,3,sobel,n.iter=50) #The example in Preacher and Hayes
```

The DV (Y) was SATIS . The IV (X) was THERAPY . The mediating variable(s) = ATTRIB .

Total Direct effect(c) of THERAPY on SATIS = 0.76 S.E. = 0.31 t direct = 2.5  
 Direct effect (c') of THERAPY on SATIS removing ATTRIB = 0.43 S.E. = 0.32 t di  
 Indirect effect (ab) of THERAPY on SATIS through ATTRIB = 0.33  
 Mean bootstrapped indirect effect = 0.31 with standard error = 0.16 Lower CI = 0.07  
 R2 of model = 0.31  
 To see the longer output, specify short = FALSE in the print statement

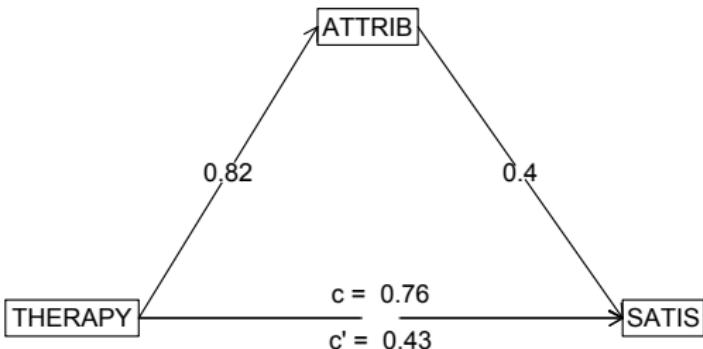
Full output

Total effect estimates (c)			
	SATIS	se	t Prob
THERAPY	0.76	0.31	2.5 0.0186
Direct effect estimates (c')			
	SATIS	se	t Prob
THERAPY	0.43	0.32	1.35 0.190
ATTRIB	0.40	0.18	2.23 0.034
'a' effect estimates			
	THERAPY	se	t Prob



## The Preacher mediation example

### Mediation model



## Take the data example from Hayes (2013)

R code

```
C.pmi <- structure(c(0.251232840197254, 0.119718779155005, 0.157470345195255,  
0.124533519925363, 0.03052112488338, 0.0734039717446355, 0.119718779155005,  
...  
33.6509729441557), .Dim = c(6L, 6L), .Dimnames = list(c("cond",  
"pmi", "import", "reaction", "gender", "age"), c("cond", "pmi",  
"import", "reaction", "gender", "age")))  
  
#n.iter set to 50 (instead of default of 5000) for speed of example  
mediate(y="reaction",x = "cond",m=c("pmi","import"),data=C.pmi,n.obs=123,n.iter=50)
```

```
Call: mediate(y = "reaction", x = "cond", m = c("pmi", "import"), data = C.pmi,  
n.obs = 123, n.iter = 50)
```

The DV (Y) was reaction . The IV (X) was cond . The mediating variable(s) = pmi import

Total Direct effect(c) of cond on reaction = 0.5 S.E. = 0.28 t direct = 1.79  
Direct effect (c') of cond on reaction removing pmi import = 0.1 S.E. = 0.24 t  
Indirect effect (ab) of cond on reaction through pmi import = 0.39  
Mean bootstrapped indirect effect = 0.7 with standard error = 0.17 Lower CI = 0.39  
R2 of model = 0.33

To see the longer output, specify short = FALSE in the print statement

Full output

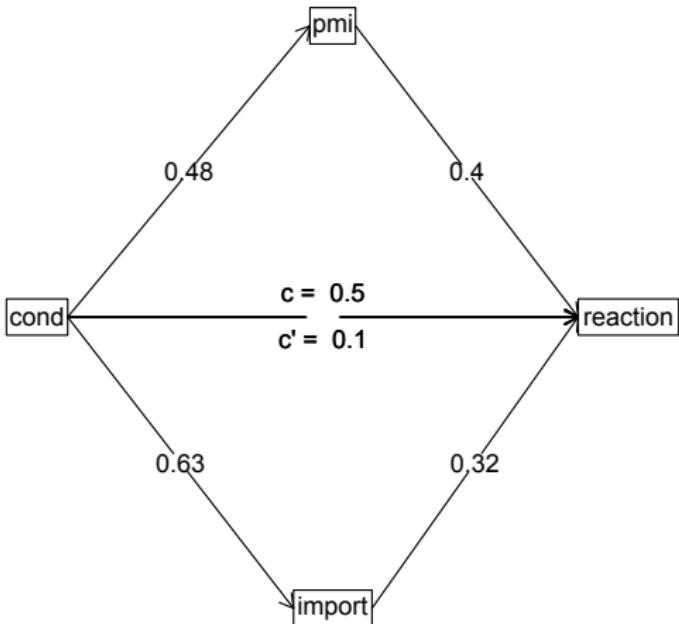
```
Total effect estimates (c)  
reaction se t Prob  
cond 0.5 0.28 1.79 0.0766
```

Direct effect estimates (c')



## The Hayes (2013) example mediation

### Mediation model



## The “New” Psychometrics

1. Classical Test theory examines responses assuming items are equivalent, or at least congeneric equivalent
2. Item Response Theory models item difficulty as well as item discrimination
3. Although seemingly very different models, factor analysis of categorical items (using tetrachoric or polychoric correlations) is equivalent to IRT 2 PL models.
4. Rasch model is just a 1 PL model where items differ in difficulty, but not discrimination.
5. 2PL has difficulty and discrimination estimated from factor analysis of tetrachoric/polychoric items.



R code

```
f1 <- irt.fa(ability)
```

```
f1 <- irt.fa(ability)
```

```
> f1
```

**Item Response Analysis using Factor Analysis**

Call: irt.fa(x = ability)

**Item Response Analysis using Factor Analysis**

**Summary information by factor and item**

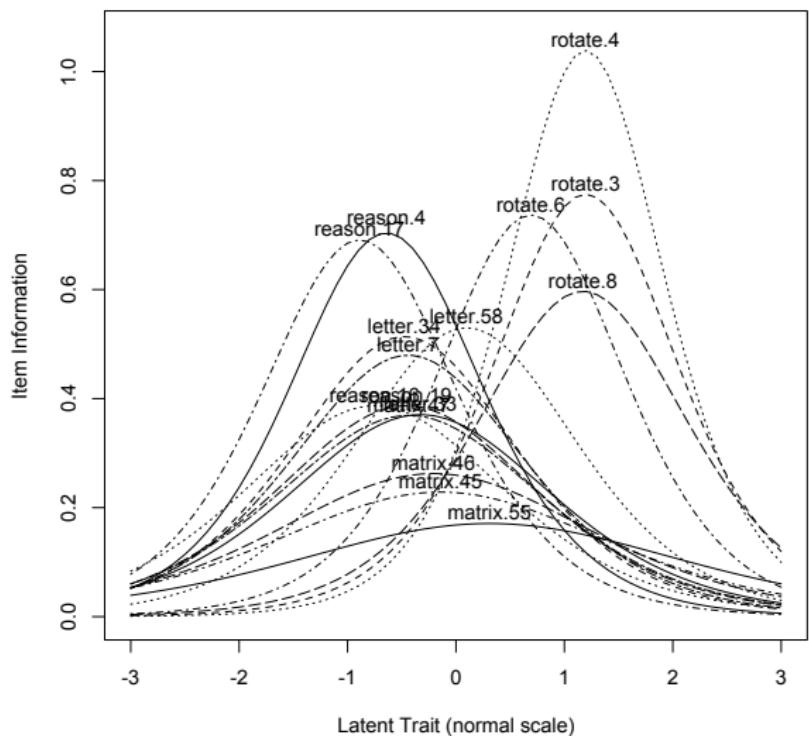
Factor = 1

	-3	-2	-1	0	1	2	3
reason.4	0.05	0.24	0.64	0.53	0.16	0.03	0.01
reason.16	0.08	0.22	0.38	0.31	0.14	0.05	0.01
reason.17	0.08	0.33	0.69	0.42	0.11	0.02	0.00
reason.19	0.06	0.17	0.35	0.36	0.19	0.07	0.02
letter.7	0.05	0.18	0.41	0.44	0.20	0.06	0.02
letter.33	0.05	0.15	0.31	0.36	0.20	0.08	0.02
letter.34	0.05	0.19	0.45	0.46	0.20	0.06	0.01
letter.58	0.02	0.09	0.30	0.53	0.35	0.12	0.03
matrix.45	0.05	0.11	0.19	0.23	0.17	0.09	0.04
...							
Test Info	0.67	2.11	4.73	5.83	5.28	2.55	0.69



## FA solution with tetrachoric correlations

Item information from factor analysis



## IRT based scoring and Classical Test Theory based scoring

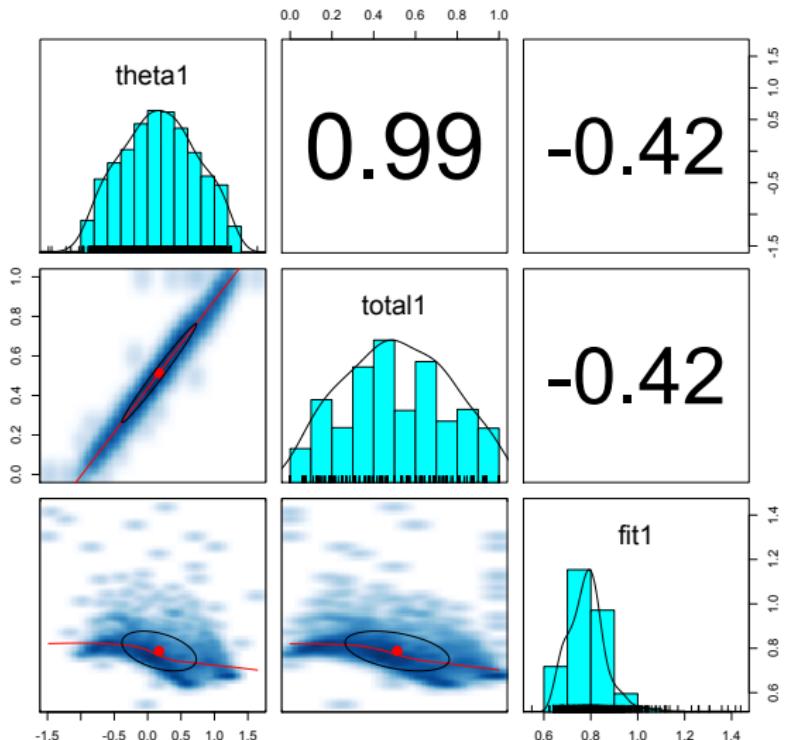
1. CTT and IRT based scores correlate almost perfectly without missing data
2. With lots of missing data, and different items for different people, IRT based scores provide more subtle distinctions.
3. `scoreIrt.2pl` and `scoreIrt.1pl` will do IRT based scores.
4. By default, will find the irt based parameters and then do the scoring.

R code

```
ability.irt <- irt.fa(ability)
ability.scores <- scoreIrt(ability.irt, ability)
pairs.panels(ability.scores, smoother=TRUE)
```



## CTT and IRT based scores are almost identical



## Multilevel reliability

- Classic reliability measures assess the variance of between person differences compared to error of the measurement.

$$\rho_{xx} = \frac{1 - \sigma_e^2}{\sigma_x^2} \quad (1)$$

- Multilevel reliability is a series of generalizability coefficients, generalizing over items, over time, time x items,

$$R_{kF} = \frac{\sigma_{id}^2 + (\sigma_{idxitems}^2/m)}{\sigma_{id}^2 + (\sigma_{idxitems}^2/m) + \sigma_{error}^2/(km)} \quad (2)$$

From Equation 6 (Shrout & Lane, 2012, p 310). See Shrout & Lane (2012) for five other generalizability formula.

- Implemented in *psych* as `mlr` or `multilevel.reliability`
- Also simulations using `sim.multi`
- I show the data from Fisher (2015) who reports 10 subjects measured over 60 (or more) days on 28 affect items.
- (Download the R data files. minor rearrangement and

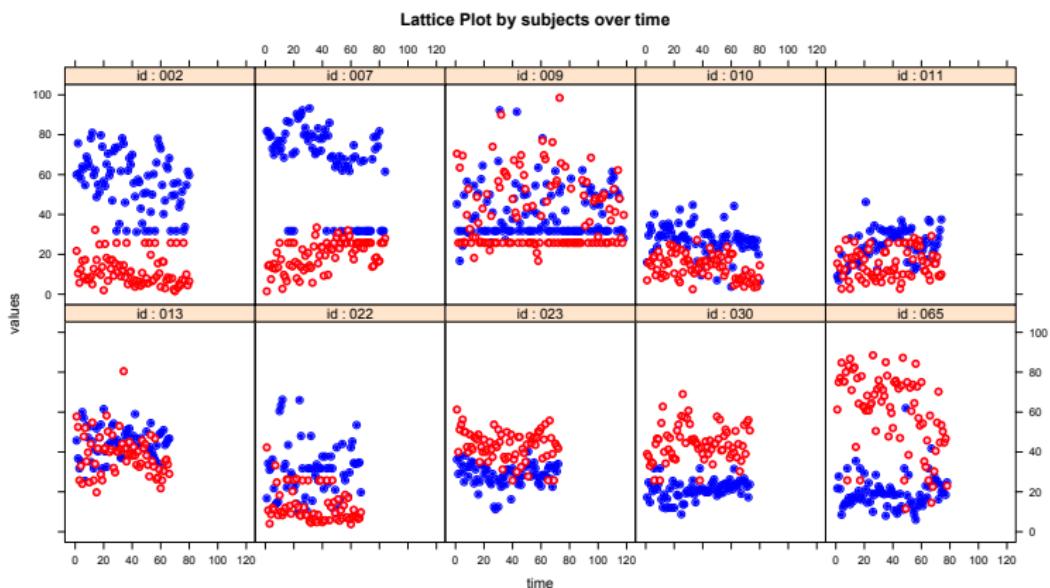


**Table:** The `multilevel.reliability` function estimates of the generalizability coefficients for the positively and negatively valenced items from Fisher (2015). RkF is the reliability of average of all ratings across all items and times (Fixed time effects), R1R is the generalizability of a single time point across all items (Random time effects), RkR is the generalizability of average time points across all items (Random time effects), Rc is the generalizability of change (fixed time points, fixed items), RkRn is the generalizability of between person differences averaged over time (time nested within people) and Rcn is the generalizability of within person variations averaged over items (time nested within people).

Multilevel reliability estimates		
Coefficient	Positive items	Negative items
RkF	1.00	1.00
R1R	0.80	0.77
RkR	1.00	1.00
Rc	0.72	0.71
RkRn	1.00	1.00
Rcn	0.64	0.59



## Assessing reliability of within subject differences in affect. Data from Fisher (2015)



## psych includes some very old ideas

1. Schmid-Leiman (Schmid & Leiman, 1957) transformations from correlated factor structures to higher order structures.
2. Dwyer extension (Dwyer, 1937; Mosier, 1938; Horn, 1973) to extend a factor solution to more variables.
3. This can be used to extend other variables into a factor space, or to relate two domains to each other.



## Extend a data set into another

First, create the data set

R code

```
set.seed(42)
d <- sim.item(12)      #two orthogonal factors
R <- cor(d)
Ro <- R[c(1,2,4,5,7,8,10,11),c(1,2,4,5,7,8,10,11)]
Roe <- R[c(1,2,4,5,7,8,10,11),c(3,6,9,12)]
fo <- fa(Ro,2)
fe <- fa.extension(Roe,fo)
fa.diagram(fo,fe=fe)
```

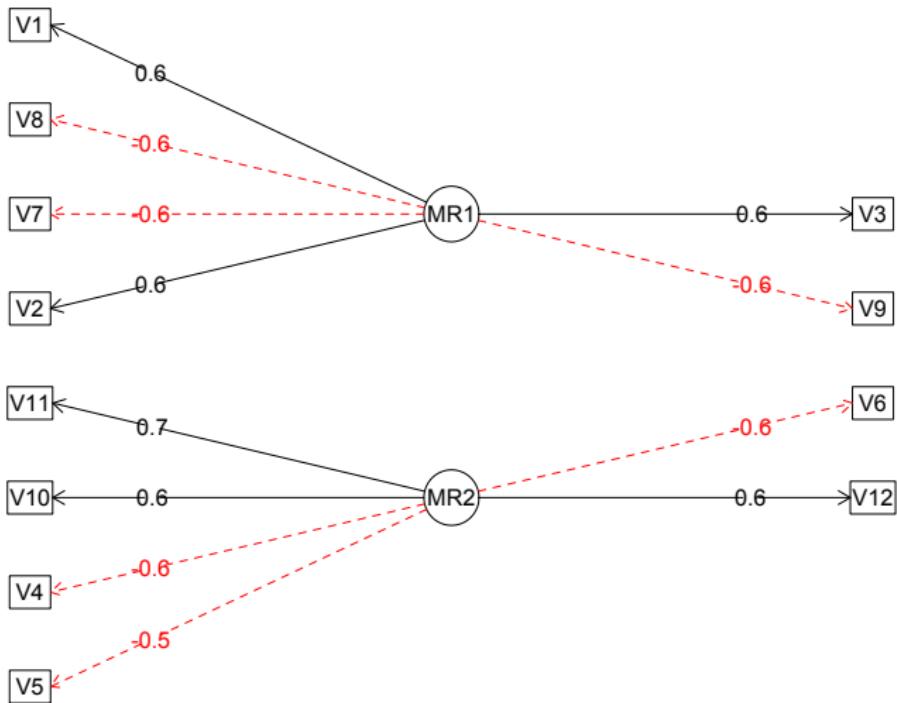
fe

```
Call: fa.extension(Roe = Roe, fo = fo)
Standardized loadings (pattern matrix) based upon correlation matrix
   MR1   MR2   h2   u2
V3  0.63 -0.02 0.39 0.61
V6  0.04 -0.61 0.37 0.63
V9 -0.61  0.01 0.38 0.62
V12 -0.06  0.58 0.33 0.67
```

	MR1	MR2
SS loadings	0.77	0.69
Proportion Var	0.19	0.17
Cumulative Var	0.19	0.37
Proportion Explained	0.53	0.47
Cumulative Proportion	0.53	1.00



## Factor extension



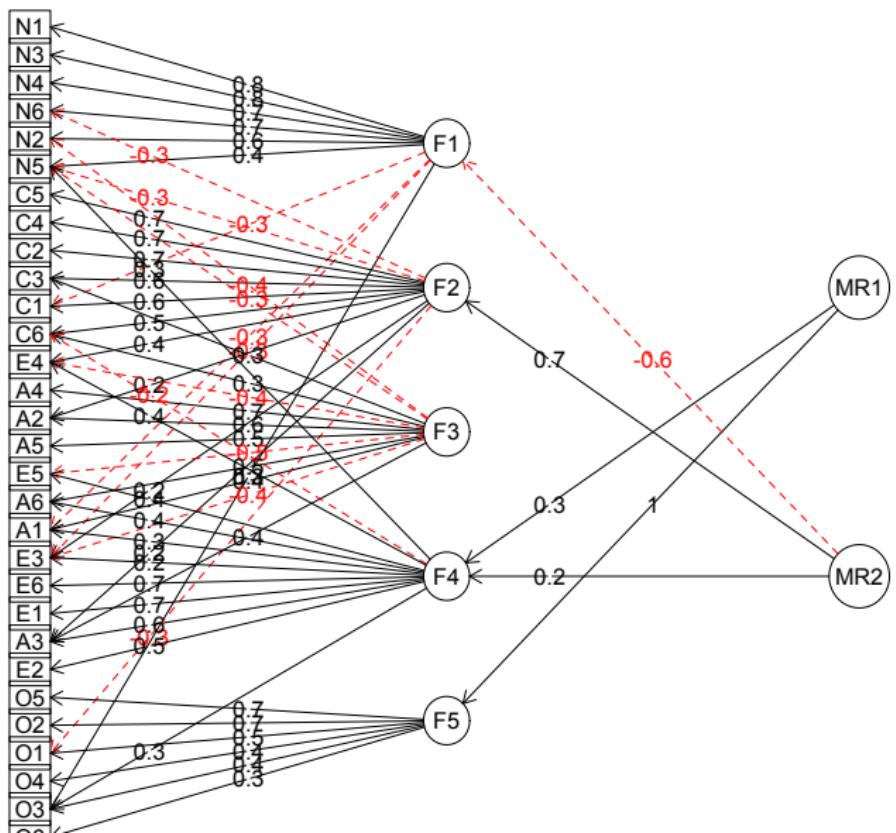
## Hierarchical factor analysis

R code

```
neo52 <- fa.multi(neo, 5, 2)  
fa.multi.diagram(neo52)
```



## Hierarchical (multilevel) Structure



ESEM can be thought of as factor extension from A to B and B to A

1. If we have two sets of variables that show factor structures within each set
  2. And then link the factor structures.
  3. This can be done in SEM, but here show how to do exploratory SEM
  4. We make up a toy data set



## R code

```
#make up a sem like problem using sim.structure
fx <-matrix(c( .9,.8,.6,rep(0,4),.6,.8,-.7),ncol=2)
fy <- matrix(c(.6,.5,.4),ncol=1)
rownames(fx) <- c("V","Q","A","nach","Anx")
rownames(fy)<- c("gpa","Pre","MA")
Phi <-matrix( c(1,0,.7,.0,1,.7,.7,.7,1),ncol=3)
gre.gpa <- sim.structural(fx,Phi,fy)
print(gre.gpa)
```

```
Call: sim.structural(fx = fx, Phi = Phi, fy = fy)
```

\$model (Population correlation matrix)

	V	Q	A	nach	Anx	gpa	Pre	MA
V	1.00	0.72	0.54	0.00	0.00	0.38	0.32	0.25
Q	0.72	1.00	0.48	0.00	0.00	0.34	0.28	0.22
A	0.54	0.48	1.00	0.48	-0.42	0.50	0.42	0.34
nach	0.00	0.00	0.48	1.00	-0.56	0.34	0.28	0.22
Anx	0.00	0.00	-0.42	-0.56	1.00	-0.29	-0.24	-0.20
gpa	0.38	0.34	0.50	0.34	-0.29	1.00	0.30	0.24
Pre	0.32	0.28	0.42	0.28	-0.24	0.30	1.00	0.20
MA	0.25	0.22	0.34	0.22	-0.20	0.24	0.20	1.00

\$reliability (population reliability)

V Q A nach Anx gpa Pre MA



## Exploratory Structural Equation Modeling

R code

```
example <- esem(gre.gpa$model, varsX=1:5, varsY=6:8, nfX=2, nfY=1,
n.obs=1000, plot=FALSE)
```

```
> example
Exploratory Structural Equation Modeling Analysis using method = minres
Call: esem(r = gre.gpa$model, varsX = 1:5, varsY = 6:8, nfX = 2, nfY = 1,
n.obs = 1000, plot = FALSE)
```

For the 'X' set:

	MR1	MR2
V	0.91	-0.06
Q	0.81	-0.05
A	0.53	0.57
nach	-0.10	0.81
Anx	0.08	-0.71

For the 'Y' set:

	MR1
gpa	0.6
Pre	0.5
MA	0.4

Correlations between the X and Y sets.

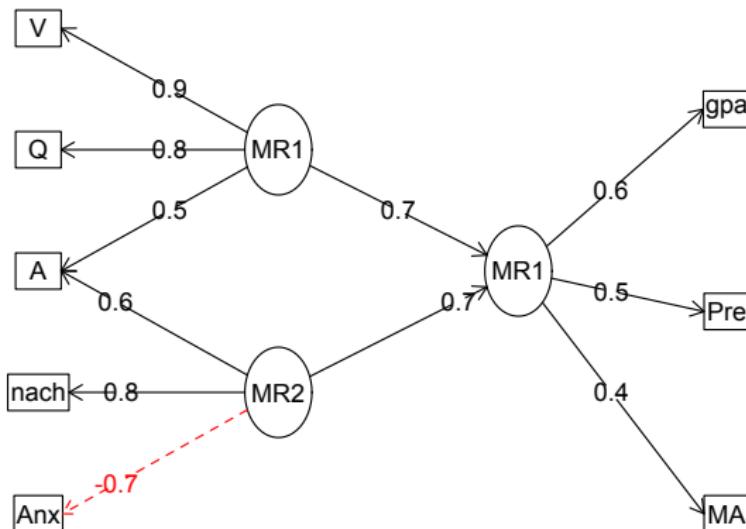
	X1	X2	Y1
X1	1.00	0.19	0.68
X2	0.19	1.00	0.67
Y1	0.68	0.67	1.00

The degrees of freedom for the null model are 56 and the empirical chi square function was 83.21  
 The degrees of freedom for the model are 7 and the empirical chi square function was 83.21



## ESEM of our toy problem

## Exploratory Structural Model



## Outline

Part I: What is R, where did it come from, why use it

- Installing R and adding packages: the building blocks of R

## Part II: A brief introduction – an overview

- R is just a fancy (very fancy) calculator
  - Descriptive data analysis
  - Some inferential analysis

### Part III R is a powerful statistical system

- Data entry (detail and practice)
  - Descriptive (again)
  - Inferential (t and F with more practice)
  - Regression
  - Basic R commands

## Part IV: Psychometrics

- Reliability and its discontents
  - EFA CFA SEM

## Part V: Help and More Help

- List of useful commands

## Part VI: The psych package and more practice



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